Tilapia viscera hydrolysate extract alleviates oxidative stress and renal damage in deoxycorticosterone acetatesalt-induced hypertension rats

by Putut Har Riyadi

Submission date: 08-Mar-2022 08:04AM (UTC+0700)

Submission ID: 1778993139

File name: C9 ARTIKEL.pdf (1.66M)

Word count: 5157

Character count: 29476

Tilapia viscera hydrolysate extract alleviates oxidative stress and renal damage in deoxycorticosterone acetate-salt-induced hypertension rats

Putut Har Riyadi¹, Mochammad Fitri Athoʻillah², Wendy Alexander Tanod^{3,5} and Irma Sarita Rahmawati⁴

 Department of Fisheries Post Harvest Technology, Faculty of Fisheries and Marine Science, Diponegoro University, Semarang 1269, Central Java, Indonesia; 2. Department of Biology, Faculty of Mathematics and Natural Science, Brawijaya University, Malang 65145, East Java, Indonesia; 3. Department of Fisheries Product Technology, Institute of Fisheries and Marine (Sekolah Tinggi Perikanan dan Kelautan), Palu 94118, Central Sulawesi, Indonesia; 4. Department of Nutrition, Faculty of Medicine, Brawijaya University, Malang 65145, East Java, Indonesia; 5. Department of Fisheries and Marine Science, Politeknik Negeri Nusa Utara, Tahuna 95821, North Sulawesi, Indonesia.

Corresponding author: Putut Har Riyadi, e-mail: putut.riyadi@live.undip.ac.id

Co-authors: MFA: athoillah.mf@gmail.com, WAT: wendytanod@stplpalu.ac.id, ISR: irmas86@gmail.com

Received: 07-06-2020, Accepted: 08-10-2020, Published online: 23-11-2020

doi: www.doi.org/10.14202/vetworld.2020.2477-2483 How to cite this article: Riyadi PH, Athoʻillah MF, Tanod WA, Rahmawati IS (2020) Tilapia viscera hydrolysate extract alleviates oxidative stress and renal damage in deoxycorticosterone acetate-salt-induced hypertension rats, Veterinary World, 13(11): 2477-2483.

Abstract

Background and Aim: Hypertension is closely related to oxidative stress conditions, which increases malondialdehyde (MDA) expression and renal damage. Tilapia viscera hydrolysate extract (TVHE) contains compounds and peptides that act as antioxidants. This study aimed to investigate TVHE therapy effect on MDA levels and renal histological conditions in deoxycorticosterone acetate (DOCA)-salt-induced hypertension rats.

Materials and Methods: Tilapia viscera were defatted and hydrolyzed using Alcalase enzyme to obtain TVHE. TVHE antioxidant activity was measured using the 1,1-diphenyl-2-picrylhydrazyl method. Fifteen Wistar male rats were divided into five groups: Normal control (without induced DOCA-salt), DOCA-salt, DOCA-salt+Captopril 5 mg/kg body weight (BW), DOCA-salt+TVHE 150 mg/kg BW, and DOCA-salt+TVHE 300 mg/kg BW. MDA level and renal histology were observed in each group.

Results: TVHE half maximal inhibitory concentration values ranged from $3.87\pm0.35~\mu g/mL$ to $42.03\pm3.55~\mu g/mL$, which were identified as in the very strong Blois category. TVHE and captopril therapy reduced MDA expression significantly (p<0.05) compared to DOCA-salt only. TVHE and captopril therapy also improved glomerular damage in DOCA-salt-induced hypertension rats.

Conclusion: TVHE has antioxidant ability, decreased MDA level, and decreased glomerular damage in DOCA-salt-induced hypertension rats.

Keywords: antioxidant, hydrolysate, peptide, tilapia, viscera.

Introduction

Hypertension is a common risk factor that is closely associated with cardiovascular and renal disease development [1]. Hypertension is a non-communicable disease that represents an increase in blood pressure and prevalence has been increasing since 2010, especially in low- to middle-income countries [2]. Approximately 1 billion people are suspected of hypertension and that number is expected to rise to 1.5 billion by 2025 [3]. Oxidative stress plays a crucial role in hypertension pathogenesis caused by the overproduction of reactive oxygen species (ROS) [4]. Animal models induced by deoxycorticosterone acetate (DOCA)-salt are a hypertensive model that is most characteristic of human cardiovascular remodeling [5].

Copyright: Riyadi, et al. Open Access. This article is distributed under the terms of the Creative Commons Attribution 4.0 International License (http://creativecommons.org/licenses/by/4.0/), which permits unrestricted use, distribution, and reproduction in any medium, provided you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons license, and indicate if changes were made. The Creative Commons Public Domain Dedication waiver (http://creativecommons.org/publicdomain/zero/1.0/) applies to the data made available in this article, unless otherwise stated.

DOCA-salt increases mineralocorticoid, followed by increased nicotinamide adenine dinucleotide phosphate (NADPH) oxidase activity and superoxide production. This process contributes to excessive oxidative stress formation and a rise in blood pressure [6]. Recent evidence indicates that hypertension is caused by oxidative stress and impaired renal function in DOCA-salt animal models [7].

Several studies have examined antioxidant and antihypertensive peptides from fish products generated by the hydrolysate technique. Products made from raw fish waste, such as chum salmon head [8], tuna fin [9], chum salmon skin [10], yellowtail tuna fin [11], Sardinela head and fins [12], tuna liver [13], skipjack egg [14], turtle egg white [15], bluefin leatherjacket head [16], bakasang skipjack [17], and peptides from fisheries waste [18], are known to possess ACE inhibitor activity. These findings indicate that fish residual waste could be a potential antihypertensive source that is nutritionally healthy, safe, cheap, and has less adverse effects due to the natural antioxidant presence [19,20]. Indonesia is the third-largest seafood consumer nation after China and Japan [21] with 91% of aquaculture production [22]. Tilapia is the third largest fish product in Indonesia after carp and salmon [23]. However, fish processing always raised environmental problems, since 50% of the waste material or residue are discarded [24]. Therefore, processing fish residues using the hydrolysis technique can add their nutraceutical and economic value.

Tilapia hydrolysate has many medicinal advantages, including high antioxidant activity in the skin [25]; the viscera suppress intracellular ROS *in vitro* [26]; the skin, bone, frame, head, muscle, and tail have high angiotensin-converting enzyme (ACE) inhibition activity [27,28]; and its viscera also reduce blood pressure, interleukin-6, and tumor necrosis factor-α expression [29]. Our preliminary study showed that tilapia hydrolysate contains adequate necessary amino acids and improved chemical characteristics, which could be met adult human nutritional needs. [30]. However, there is still a lack of information about the tilapia viscera hydrolysate extract (TVHE) effect on oxidative stress and renal damage in hypertensive rats induced by DOCA-salt.

This study investigated the TVHE antioxidant potential on malondialdehyde (MDA) levels and renal histological conditions. This study used DOCA-salt-induced hypertensive rat models to determine the effectiveness of TVHE therapy as a nutraceutical candidate.

Materials and Methods

Ethical approval

Animal experiments approved by the Research Ethics Committee, Brawijaya University, Indonesia (Ref. No. 1064-Kep-UB).

Study period and location

The research was conducted from April to September 2019 at the Experimental Animal Laboratory, Institute of Bioscience, Brawijaya University, Indonesia.

Sample preparation and extraction

Tilapia viscera were obtained from PT Aquafarm Nusantara, Semarang Industrial Estate, in Indonesia. Viscera were cleaned with water and defatted. The hydrolysis process was carried out using the Alcalase enzyme (Sigma-Aldrich No. 126741) so that TVHE rich in bioactive peptides was obtained. TVHE was optimized using the response surface methodology (RSM) to obtain TVHE with the best hydrolysis degree [31].

Antioxidant assay

Antioxidant activity was measured using the 1,1-diphenyl-2-picrylhydrazyl (DPPH, Merck) radical scavenging method [32,33]. Twenty-five milligrams of TVHE were added to methanol at an extract concentration of 200 μ g/mL. The half maximal inhibitory concentration (IC ₅₀) was determined using 30, 60, 90, 120, and 150 μ g/mL TVHE concentrations. Ascorbic acid (Vitamin C) was used as a comparison control. Two milliliters of each extract solution

were added to 2 mL of 50 μM DPPH solution. The solution was homogenized and left for 30 min in a dark room before measuring with a spectrophotometer (Shimadzu UV-1800) at 517 nm absorbance. The DPPH solution absorbance value was also measured as a blank. The DPPH radical scavenging assay was performed in three independent experiments, and the measurement results were expressed as the mean±standard deviation using Microsoft Excel 2016.

The DPPH radical scavenging effect calculated using the equation:

% DPPH Scavenging Effect =

 $\frac{\text{Blank Absorbance} - \text{Sampel Absorbance}}{\text{Blank Absorbance}} \times 100\%$

Animal subject preparations

TVHEs with the best hydrolysis degree were used for therapy in hypertensive Wistar rats induced by DOCA-salt. Fifteen male rats (200-210 g) were adapted for 14 days in a controlled room with 24±2°C, humidity 50-60%, and 12 h dark/light cycle. Rats were given mineral water and standard food (15% crude protein, 12% water content, crude fiber up to 6%, crude fat 3-7%, maximum ash 7%, calcium 0.9-1.1%, and phosphorus 0.6-0.9%). In the pre-treatment phase, DOCA-salt was dissolved in corn oil and orally administered through gavage (except in control rats) 10 times for 5 weeks (2 times a week) to induce hypertension. Two DOCA-salt concentrations were prepared: 20 mg/kg and 10 mg/kg DOCA-salt. At the end of the 5th week, the rats were measured for blood pressure, then divided into five treatment groups: Normal control (without induced DOCAsalt), DOCA-salt, DOCA-salt+Captopril 5 mg/kg of body weight (BW), DOCA-salt+TVHE 150 mg/kg of BW, and DOCA-salt+TVHE 300 mg/kg of BW. Rats were given TVHE and captopril treatments as therapy for 8 days orally through gavage. Systolic blood pressure (SBP) was measured by the tail-cuff method using a blood pressure analyzer. Rats were considered as hypertensive if the SBP was equal to or higher than 150 mmHg [34].

MDA measurement

Rats were anesthetized with ketamine at the dose of 70 mg/kg BW and dissected to collect the kidney [35]. Kidneys were cut into small pieces and then crushed in a cold mortar on ice. Then, 1 ml of physiological NaCl added. The homogenate was centrifuged at 8000 rpm for 20 min, and the supernatant was taken. The supernatant (100 μ L) was diluted by adding 450 μ L of distilled water, followed by adding 100 μ L trichloroacetic 100%, 250 μ L HCl 1 N, and 100 μ L Na Thio 1%. The mixture was homogenized using a vortex and centrifuged at 5000 rpm for 15 min. The supernatant was transferred to a new microcentrifuge tube and incubated in a water bath at 100°C for 10 min. The absorbance was measured with a spectrophotometer (Shimadzu UV-1800) at a wavelength of

532.8 nm. After the absorbance data were obtained, the concentrations were calculated using a standard curve.

Histopathological analysis

Renal tissue histology was carried out according to routine laboratory procedures. After DOCA-salt induction and TVHE therapy, rats were sacrificed using ketamine at the dose of 70 mg/kg BW, and the kidney was taken and fixed with 10% formaldehyde solution for 24 h. The pathological structure was observed by making paraffin renal tissue. After hematoxylin-eosin staining, renal histopathological examination was visualized using an Olympus XC10 camera with a 400×. Renal histopathological tissue images were analyzed using the Olympus Viewer for Imaging Applications program to see cell changes in the renal glomerular and tubular portions.

Statistical analysis

MDA data are expressed as mean±standard deviation. The Statistical Package for the Social Sciences version 20.0 (IBM Corp., Armonk, NY) was used to analyze the data. All data sets were tested for normal distributions using the Kolmogorov–Smirnov test and were normally distributed. The difference between the groups was analyzed using a one-way analysis of variance followed by the Duncan *post hoc* test. p<0.05 was considered statistically significant.

Results

Antioxidant activity of tilapia viscera hydrolyzed extract

The optimization results using the RSM method [31] were obtained for the four TVHEs with the highest degree of hydrolysis, 34.98%, 35.55%, 36.01%, and 41.46%, respectively. The four TVHEs with the highest percentage of hydrolysis were tested for DPPH radical scavenging activity. The IC_{50} results for the four TVHEs are shown in Table-1. The results showed that increased hydrolysis in the TVHEs increased the radical DPPH scavenging percentage in a dose-dependent manner (Figure-1).

Therapeutic effect of tilapia viscera TVHE on MDA levels

This study demonstrated that DOCA-salt administration in rats significantly increased the concentration of MDA (p<0.05) compared to normal rats (Figure-2). Both administrations of TVHE at 150 and

 $\textbf{Table-1:} \ \text{IC}_{\text{50}}$ of tilapia viscera TVHE using DPPH radical scavenging method.

Degree of hydrolysis (%)	Mean of IC _{so} (μg/mL)	
34.98	42.03±3.55	
35.55	13.03±2.28	
36.01	8.99±3.36	
41.46	3.87±0.35	
Ascorbic acid	1.53±0.36	

IC₅₀=Half maximal inhibitory concentration, TVHE=Tilapia viscera hydrolysate extract, DPPH=1,1-diphenyl-2-picrylhydrazyl 300 mg/kg of BW significantly reduced the MDA concentration (p<0.05) in DOCA-salt-induced hypertensive rats compared to the DOCA-salt control group. However, these MDA levels were not fully reduced to the normal group MDA levels. Interestingly, there was no substantial difference in MDA concentration in rats treated with captopril and TVHE. These data show that TVHE can induce a therapeutic effect on lower MDA concentrations in DOCA-salt-induced hypertensive rats.

Therapeutic TVHE effect on renal histopathology

Renal histopathology demonstrated that there were distinct and clear pathological changes, such as necrosis in the glomerular area surrounded by Bowman capsules and renal tubules in DOCA-salt-induced rats compared to normal rat (Figure-3a and b). However, the renal pathology improved in DOCA-salt rats after treated with TVHE (150 and 300 mg/kg of BW) or captopril (5 mg/kg of BW) (Figure-3c-e). The histopathological results showed that TVHE promotes repair in renal pathologies in DOCA-salt-induced hypertension rats.

Discussion

Using fish by-products processed by the hydrolysis technique have been highlighted in the present year due to their beneficial effects on health. Some of the hydrolysated fish by-products that can act as an antihypertensive are salmon heads, skate skin (Okamejeri kenojei), salmon skin, gelatin from thornback ray skin, smooth-hound viscera (Mustelus mustelus), tuna fins, meat red tuna, tuna liver, skipjack egg, round Sardinella head and viscera (Sardinella aurita), and yellowfin sole bone (Limanda aspera) [18]. Our present study demonstrated that TVHE has a strong antioxidant capacity and has beneficial effects on lowering MDA due to increased oxidative stress. This ultimately leads to glomerular repair in hypertensive rats induced by DOCA-salt.

The DPPH antioxidant assay is the most popular tool to evaluate a substance's ability to inhibit free radicals because it is simple, sensitive, and a fast way to test antioxidants [36,37]. The DPPH reduction measured in the TVHE samples was proportional to the amount of purple solution turning yellow to calculate an IC_{50} value, suggesting that 50% of the DPPHfree radicals are scavenged in solution [33]. Table-1 shows the IC₅₀ for the four TVHEs. Blois [38] categorized antioxidant activity: Weak (IC50 ranges from 150 to 200 μg/mL); moderate (IC₅₀ ranges from 100 to 150 μ g/mL); strong (IC₅₀ between 50 and 100 μ g/mL); and very strong (IC_{so} <50 μg/mL). Based on the Blois category, the TVHE antioxidant ability was very strong. TVHE was almost comparable to ascorbic acid (Vitamin C), an antioxidant substance that is widely used. A previous study reported Vitamin C IC₅₀ values ranging from 1.01 to 58.94 µg/mL, depending on the Vitamin C concentration used [32].

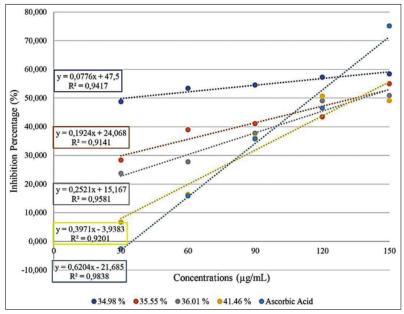


Figure-1: Percentage of 1,1-diphenyl-2-picrylhydrazyl radical inhibition of tilapia viscera hydrolysate extract and ascorbic acid at various concentrations.

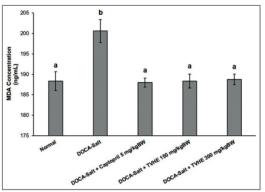


Figure-2: Malondialdehyde concentration in rats treated with deoxycorticosterone acetate (DOCA)-salt, DOCA-salt, and tilapia viscera hydrolysate extract. The values are represented as mean±SD (n=3 for each group). Different letters on the figure considered significantly different for each group at p<0.05 and vice versa based on the Duncan post hoc test.

Table-1 also shows that the hydrolysis degree was directly proportional to the TVHE IC_{50} . This suggests the hydrolysis process increased the peptide number [31,39]. These peptides tend to be short-chain peptides. Simple short-chain peptides can scavenge free radicals with ease [40]. This work suggests the hydrolysis process breaks down TVHE proteins susceptible to oxidation and releasing new peptides in a specific manner. These peptides can be further broken down using enzymes, acids, or bases in the body [41]. This would be advantageous, as shorter and simpler peptide chains have higher antihypertensive

potential as bioactive peptides, reducing oxidative stress [29,42].

The hypertension condition causes oxidative stress and produces excess free radicals (ROS) [4]. DOCA-salt-induced rats have endocrine hypertension, which increases oxidative stress [5]. These free radicals increase lipid peroxidation, which breaks down into MDA. MDA is a cellular defect marker caused by free radicals [31,43,44]. Figure-2 shows that DOCA-salt induction caused an increase in MDA expression in rats. DOCA-salt induction increases free radicals, which liberating renal phospholipids. These lipids are further broken down by peroxide to produce MDA [33,45].

Figure-2 also shows that DOCA-salt induction resulted in a significant increase in MDA levels (6.57%). The MDA level increase was related to ROS, was correlated with an increase in blood pressure. NADPH oxidase activation is known to be elevated in the aortic cell membrane at this time, producing ROS in the form of anion superoxide (${\rm O_2}^-$) [34]. Increased ROS affects lipid peroxidation and the final product, MDA [35]. Our study suggests that TVHE and captopril therapy reduce MDA levels by 5.90-6.27%. Our result suggests a beneficial effect for captopril and TVHE therapies by reducing the blood pressure and MDA levels in hypertensive rats.

TVHE and captopril may reduce MDA levels by inhibiting ACE, angiotensin-I and angiotensin-II, further reducing aldosterone secretion. Low aldosterone secretion results in decreased blood pressure, NADPH oxidase inactivation, low ROS, and decreased MDA [46]. Our study showed TVHEs ability to reduce antioxidant activity by scavenging free

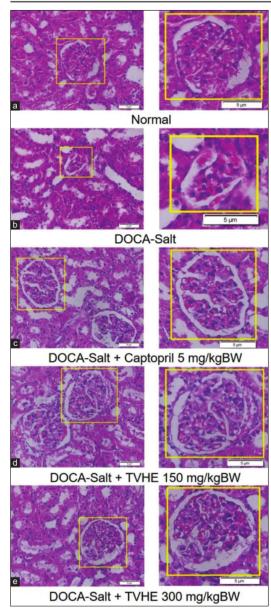


Figure-3: (a-e) Tilapia viscera hydrolysate extract repairs histopathological changes in renal tissues in deoxycorticosterone acetate-salt rats. Renal was stained with hematoxylin and eosin and observed at $400\times$.

radicals and reducing MDA levels. A previous study treated DOCA-salt rats with 200 mg/kg of bakasang for a week and also reduced MDA levels. Bakasang is a traditional food from Maluku and North Sulawesi made from fermented skipjack tuna [47]. These results showed that fish products have a beneficial effect on reducing MDA levels in DOCA-salt hypertensive rats.

Figure-3 shows the glomerular area surrounded by the Bowman capsules and renal tubules are

necrotic in DOCA-salt rats. The necrosis caused the presence of karyolysis in cell nuclei. This is caused by DOCA-salt-induced hypertension, oxidative stress, and subsequent renal damage [35,48]. The TVHE and captopril therapies improved the glomerular tissue structure close to what was observed in normal rats.

TVHE therapy showed antioxidant ability by inhibiting lipid peroxidation, which would further increase enzymatic antioxidants, such as superoxide dismutase in the kidneys. Increased enzymatic antioxidants could decrease the free radical anion superoxide (O_2^{-}) [38,49]. Often, endogenous antioxidants and exogenous antioxidants work together to properly inhibit excessive ROS formation [39,40]. TVHE acts as an exogenous antioxidant that can work by directly donating hydrogen ions to neutralize free radicals' toxic effects. In addition, TVHE can work indirectly by increasing the endogenous antioxidant expression genes through several mechanisms in the body [50,51].

Conclusion

TVHE may be an antioxidant that inhibits MDA levels as a consequence of increased oxidative stress in DOCA-salt-induced hypertensive rats. TVHE also demonstrates the ability to restore glomerular tissue damaged due to hypertension in renal tissue histology. However, further investigation is required to assess TVHE bioavailability and efficacy after digestion.

Authors' Contributions

PHR has made a significant contribution to conception, design, interpretation of data, drafting, revising the manuscript, and gave final approval of the version to be published. MFA helped in data collection, edited article, and made critical revisions. WAT made a substantial contribution to the acquisition of data, analysis, and drafting of the manuscript. ISR analyzed data, drafted the article, and made a critical revision. All authors read and approved the final manuscript.

Acknowledgments

We want to acknowledge the Indonesia Endowment Fund for Education (LPDP-BUDI DN), the Ministry of Finance, and the Ministry of Education and Culture, the Republic of Indonesia, for offering a scholarship to the first author for his research with grant no. PRJ-6468/LPDP.3/2016.

Competing Interests

The authors declare that they have no competing interests.

Publisher's Note

Veterinary World remains neutral with regard to jurisdictional claims in published institutional affiliation.

References

 Mills, K.T., Chen, J., Yang, W., Appel, L.J., Kusek, J.W., Alper, A., Delafontaine, P., Keane, M.G., Mohler, E.,

- Ojo, A., Rahman, M., Ricardo, A.C., Soliman, E.Z., Steigerwalt, S., Townsend, R. and He, J. (2016) Sodium excretion and the risk of cardiovascular disease in patients with chronic kidney disease. *JAMA*, 315(20): 2200.
- Mills, K.T., Stefanescu, A. and He, J. (2020) The global epidemiology of hypertension. *Nat. Rev. Nephrol.*, 16(4): 223-237.
- Guilbert, J.J. (2003) The World Health Report 2002-reducing Risks, Promoting Healthy Life. Vol. 16. Education for Health, Change in Learning and Practice, United Kingdom. p. 30-230.
- Pawluk, H., Pawluk, R., Robaczewska, J., Kędziora-Kornatowska, K. and Kędziora, J. (2017) Biomarkers of antioxidant status and lipid peroxidation in elderly patients with hypertension. *Redox Rep.*, 22(6): 542-546.
- Lin, H.Y., Lee, Y.T., Chan, Y.W. and Tse, G. (2016) Animal models for the study of primary and secondary hypertension in humans (review). *Biomed. Rep.*, 5(6): 653-659.
- Beswick, R.A., Dorrance, A.M., Leite, R. and Webb, R.C. (2001) NADH/NADPH oxidase and enhanced superoxide production in the mineralocorticoid hypertensive rat. *Hypertension*, 38(5): 1107-1111.
- Seifi, B., Kadkhodaee, M., Karimian, S.M., Zahmatkesh, M., Xu, J. and Soleimani, M. (2010) Evaluation of renal oxidative stress in the development of DOCA-salt-induced hypertension and its renal damage. Clin. Exp. Hypertens., 32(2): 90-97.
- Ohta, T., Iwashita, A., Sasaki, S. and Kawamura, Y. (1997) Antihypertensive action of the orally administered protease hydrolysates of chum salmon head and their angiotensin I-converting enzyme inhibitory peptides. Food Sci. Technol. Tokyo, 3(4): 339-343.
- Lee, S.H., Qian, Z.J. and Kim, S.K. (2010) A novel angiotensin I converting enzyme inhibitory peptide from tuna frame protein hydrolysate and its antihypertensive effect in spontaneously hypertensive rats. Food Chem., 118(1): 96-102.
- Lee, J.K., Jeon, J.K. and Byun, H.G. (2014) Antihypertensive effect of novel angiotensin I converting enzyme inhibitory peptide from chum salmon (*Oncorhynchus keta*) skin in spontaneously hypertensive rats. J. Funct. Foods, 7(2): 381-389
- Jung, W.K., Mendis, E., Je, J.Y., Park, P.J., Son, B.Y., Kim, H.C., Choi, Y.K. and Kim, S.K. (2006) Angiotensin I-converting enzyme inhibitory peptide from yellowfin sole (*Limanda aspera*) frame protein and its antihypertensive effect in spontaneously hypertensive rats. Food Chem., 94(1): 26-32.
- Bougatef, A., Nedjar-Arroume, N., Ravallec-Plé, R., Leroy, Y., Guillochon, D., Barkia, B. and Nasri, M. (2008) Angiotensin I-converting enzyme (ACE) inhibitory activities of sardinelle (Sardinella aurita) by-products protein hydrolysates obtained by treatment with microbial and visceral fish serine proteases. Food Chem., 111(2): 350-356.
- Je, J.Y., Lee, K.H., Lee, M.H. and Ahn, C.B. (2009) Antioxidant and antihypertensive protein hydrolysates produced from tuna liver by enzymatic hydrolysis. *Food Res. Int.*, 42(9): 1266-1272.
- Intarasirisawat, R., Benjakul, S., Wu, J. and Visessanguan, W. (2013) Isolation of antioxidative and ACE inhibitory peptides from protein hydrolysate of skipjack (*Katsuwana pelamis*) roe. *J. Funct. Foods*, 5(4): 1854-1862.
- Rawendra, R.D.S., Aisha, A., Chang, C.I., Aulanni'am, A., Chen, H.H., Huang, T.C. and Hsu, J.K. (2013) A novel angiotensin-converting enzyme inhibitory peptide derived from proteolytic digest of Chinese soft-shelled turtle egg white proteins. J. Proteomics, 94(17): 359-369.
- Chi, C.F., Wang, B., Wang, Y.M., Zhang, B. and Deng, S.G. (2015) Isolation and characterization of three antioxidant peptides from protein hydrolysate of bluefin leatherjacket (Navodon septentrionalis) heads. J. Funct. Foods, 12(1): 1-10.

- Wenno, M.R., Suprayitno, E., Aulanni'am, A. and Hardoko, H. (2016) Identification and molecular interaction mechanism angiotensin-converting enzyme inhibitory peptide from bakasang (fermented skipjack tuna (*Katsuwonus pelamis*)). *Int. J. PharmTech Res.*, 9(12): 591-598.
- Riyadi, P.H. (2018) Bioactive peptide for lowering pressure blood from fisheries by-product: A review. J. Peng. Biotek. Hasil. Pi., 7(1): 1-6.
- Samaranayaka, A.G.P. and Li-Chan, E.C.Y. (2011) Foodderived peptidic antioxidants: A review of their production, assessment, and potential applications. *J. Funct. Foods*, 3(4): 229-254.
- Sila, A. and Bougatef, A. (2016) Antioxidant peptides from marine by-products: Isolation, identification and application in food systems. A review. J. Funct. Foods, 21(2): 10-26
- Guillen, J., Natale, F., Carvalho, N., Casey, J., Hofherr, J., Druon, J.N., Fiore, G., Gibin, M., Zanzi, A. and Martinsohn, J.T. (2019) Global seafood consumption footprint. *Ambio*, 48(2): 111-122.
- Belton, B., Bush, S.R. and Little, D.C. (2018) Not just for the wealthy: Rethinking farmed fish consumption in the global South. *Global Food Secur.*, 16(1): 85-92.
- Indonesia: Production Volume Tilapia. (2017) Statista. Available from: https://www.statista.com/statistics/1083241/indonesia-production-volume-of-tilapia. Retrieved on 10-09-2020.
- Villamil, O., Váquiro, H. and Solanilla, J.F. (2017) Fish viscera protein hydrolysates: Production, potential applications and functional and bioactive properties. Food Chem., 224(10): 160-171.
- Zhang, Y., Duan, X. and Zhuang, Y. (2012) Purification and characterization of novel antioxidant peptides from enzymatic hydrolysates of tilapia (*Oreochromis niloticus*) skin gelatin. *Peptides*, 38(1): 13-21.
- Gómez, L.J., Gómez, N.A., Zapata, J.E., López-García, G., Cilla, A. and Alegría, A. (2019) *In-vitro* antioxidant capacity and cytoprotective/cytotoxic effects upon Caco-2 cells of red tilapia (*Oreochromis* spp.) viscera hydrolysates. *Food Res. Int.*, 120(6): 52-61.
- Roslan, J., Faezah, K., Abdullah, N. and Mazlina, S. (2014) Characterization of fish protein hydrolysate from tilapia (Oreochromis niloticus) by-product. Agric. Agric. Sci. Proc., 2(1): 312-319.
- Yesmine, B.H., Antoine, B., da Silva Ortência Leocádia, N.G., Rogério, B.W., Ingrid, A., Nicolas, B., Thierry, M., Jean-Marie, P., Frédéric, S. and Stéphanie, B.J. (2017) Identification of ace inhibitory cryptides in tilapia protein hydrolysate by UPLC-MS/MS coupled to database analysis. J. Chromatogr. B Analyt. Technol. Biomed. Life Sci., 1052(10): 43-50.
- Riyadi, P.H., Tanod, W.A., Sulistiyati, D.T., Aulanni'am, A. and Suprayitno, E. (2020) Effects of nile tilapia (Oreochromis niloticus) viscera hydrolyzate on blood pressure, TNF-a and IL-6 expression in rats (Rattus norvegicus) induced by DOCA-salt. Indian J. Anim. Res., https://www.arccjournals.com/journal/indian-journal-of-animal-research/B-1195?type=onlineFirstArticle. Retrieved on 10-11-2020.
- Riyadi, P.H., Suprayitno, E., Aulanni'am, A., and Sulistiyati, T.D. (2019) Chemical characteristics and amino acids profile of protein hydrolysates of nile tilapia (Oreochromis niloticus) Viscera. J. Worlds Poult. Res., 9(4): 324-328.
- Riyadi, P.H., Suprayitno, E., Aulanni'am, A., and Sulistiati, T.D. (2019) Optimization of protein hydrolysate from visceral waste of nile tilapia (*Oreochromis niloticus*) by response surface methodology. *AACL Bioflux*, 12(6): 2347-2358.
- Dewanto, D.K., Finarti, F., Hermawan, R., Ndobe, S., Riyadi, P.H. and Tanod, W.A. (2019) Aktivitas antioksidan ekstrak karang lunak asal teluk palu, Sulawesi Tengah,

- Indonesia. J. Pascapanen Bioteknol. Kelautan Perikanan., 14(2): 163.
- Tanod, W.A., Dewanto, D.K., Ndobe, S., Riyadi, P.H. and Putra, M.A. (2019) Screening of antibacterial and antioxidant activity from the soft Corals Sinularia sp. and Sarcophyton sp. Origin palu bay, Central Sulawesi, Indonesia. Squalen Bull. Mar. Fish. Postharvest Biotech., 14(2): 73-83.
- Kloza, M., Baranowska-Kuczko, M., Malinowska, B., Karpińska, O., Harasim-Symbor, E., Kasacka, I. and Kozłowska, H. (2017) The influence of DOCA-salt hypertension and chronic administration of the FAAH inhibitor URB597 on KCa2.3/KCa3.1-EDH-type relaxation in rat small mesenteric arteries. Vasc. Pharmacol., 99(8): 65-73.
- Aulanni'am, A., Roosdiana, A. and Rahmah, N.L. (2012)
 The potency of Sargassum duplicatum bory extract on inflammatory bowel disease therapy in Rattus norvegicus. J. Life Sci., 6(2): 144-154.
- Atho'illah, M.F., Safitri, Y.D., Nur'aini, F.D., Savitri, R.U., Rahayu, S., Widyarti, S. and Rifa'il, M. (2019) Evaluation of glyceollin accumulation and antioxidant properties on soybean (Glycine max L.) through combination of different biotic elicitor and light. Sci. Study Res. Chem. Chem. Eng. Biotechnol. Food Ind., 20(2): 199-208.
- Tanod, W.A, Yanuhar, U., Maftuch, M., Wahyudi, D.S. and Risjani, Y. (2019) DPPH scavenging property of bioactives from soft corals origin Palu Bay, Central Sulawesi, Indonesia. *IOP Conf. Ser. Earth Environ. Sci.*, 236(16): 012121.
- Blois, M.S. (1958) Antioxidant determinations by the use of a stable free radical. *Nature*, 181: 1199-1200.
- Riyadi, P.H., Wahyudi, D. and Tanod, W.A. (2019) Effects
 of dichloromethane Sarcophyton spp. Extract on the lipopolysaccharide-induced expression of nuclear factor-kappa
 B and inducible nitric oxide synthase in mice. Vet. World,
 12(12): 1897-1902.
- Zou, T.B., He, T.P., Li, H.B., Tang, H.W. and Xia, E.Q. (2016) The structure-activity relationship of the antioxidant peptides from natural proteins. *Molecules*, 21(1): 72.
- Kim, S.K. and Wijesekara, I. (2010) Development and biological activities of marine-derived bioactive peptides: A

- review. J. Funct. Foods, 2(1): 1-9.
- Li, Y. and Yu, J. (2015) Research progress in structure-activity relationship of bioactive peptides. *J. Med. Food*, 18(2): 147-156.
- Gofur, A., Witjoro, A., Ningtiyas, E.W., Setyowati, E., Mukharromah, S.A., Atho'illah, M.F. and Lestari, S.R. (2019) The evaluation of dietary black soybean and purple sweet potato on insulin sensitivity in streptozotocin-induced diabetic rats. *Pharmacogn. J.*, 11(4): 639-646.
- Rahmawati, I.S. (2018) Malonaldehyde level of administration ethanol extract of purple sweet potato var. Ayamurasaki in DOCA-salt hypertensive rats. J. Appl. Food Technol., 5(1): 6-9.
- Chakrabarti, S., Jahandideh, F. and Jianping, W. (2014) Food-derived bioactive peptides on inflammation and oxidative stress. *Biomed Res. Int.*, 2014: 1-11.
- Safaeian, L., Emami, R., Hajhashemi, V. and Haghighatian, Z. (2018) Antihypertensive and antioxidant effects of protocatechuic acid in deoxycorticosterone acetate-salt hypertensive rats. *Biomed. Pharmacother.*, 100(4): 147-155.
- Nurmahdi, H., Prasetyawan, S., Wenno, M.R. and Aulanni'am, A. (2017) Antihypertension effect from Bakasang's peptide extract based on MDA levels in sera and iNOS expression in cardiac tissue of rats hypertensive model. *Int. J. Pharm. Clin. Res.*, 9(2): 129-134.
- Noori, S. and Mahboob, T. (2010) Antioxidant effect of carnosine pretreatment on cisplatin-induced renal oxidative stress in rats. *Indian J. Clin. Biochem.*, 25(1): 86-91.
- Iyer, A., Chan, V. and Brown, L. (2010) The DOCA-salt hypertensive rat as a model of cardiovascular oxidative and inflammatory stress. *Curr. Cardiol. Rev.*, 6(4): 291-297.
- Lan, C., Ding, L. and Su, Y. (2015) Grape seed proanthocyanidins prevent DOCA-salt hypertension-induced renal injury and its mechanisms in rats. *Food Funct.*, 6(7): 2179-2186.
- Tkaczewski, W., Kedziora, J., Buczyński, A., Dziekański, S. and Ryniec, A. (1989) Effect of captopril on superoxide dismutase (SOD-1) activity and malondialdehyde (MDA) level in blood platelets in patients with arterial hypertension. Kardiol. Pol., 32(3): 138-141.

Tilapia viscera hydrolysate extract alleviates oxidative stress and renal damage in deoxycorticosterone acetate-salt-induced hypertension rats

ORIGINALITY REPORT

16% SIMILARITY INDEX

17%
INTERNET SOURCES

7%
PUBLICATIONS

5% STUDENT PAPERS

MATCH ALL SOURCES (ONLY SELECTED SOURCE PRINTED)

10%

★ doaj.org

Internet Source

Exclude quotes On

Exclude bibliography On

Exclude matches

< 3%

Tilapia viscera hydrolysate extract alleviates oxidative stress and renal damage in deoxycorticosterone acetate-salt-induced hypertension rats

GRADEMARK REPORT	
FINAL GRADE	GENERAL COMMENTS
/0	Instructor
PAGE 1	
PAGE 2	
PAGE 3	
PAGE 4	
PAGE 5	
PAGE 6	
PAGE 7	