

The increasing of superoxide dismutase (SOD) in metabolic syndrome rats by robusta coffee leaves (*Coffea canephora*)

by Gemala Anjani

Submission date: 05-Dec-2022 08:47AM (UTC+0700)

Submission ID: 1971314531

File name: ing_of_superoxide_dismutase_SOD_in_metabolic_-_Gemala_Anjani.pdf (326.61K)

Word count: 5564

Character count: 28897

The increasing of superoxide dismutase (SOD) in metabolic syndrome rats by robusta coffee leaves (*Coffea canephora*)

¹Almira, V.G., ^{1,2*}Widyastuti, N., ^{1,2}Anjani, G., ³Widyastiti, N.S., ^{1,2}Panunggal, B.,
^{1,2}Nissa, C., ^{1,2}Tsani, A.F.A., ^{1,2}Wijayanti, H.S. and ^{1,2}Purwanti, R.

¹Department of Nutrition Science, Faculty of Medicine, Universitas Diponegoro, Jl. Prof H. Soedarto, SH, Tembalang, Semarang 50275, Indonesia

²Center of Nutrition Research (CENURE), Universitas Diponegoro, Jl. Prof H. Soedarto, SH, Tembalang, Semarang 50275, Indonesia

³Department of Clinical Pathology, Faculty of Medicine, Universitas Diponegoro, Jl. Prof H. Soedarto, SH, Tembalang, Semarang 50275, Indonesia

Article history:

Received: 29 December 2019

Received in revised form: 4 February 2020

Accepted: 6 February 2020

Available Online: 30 May 2020

Keywords:

Superoxide dismutase,
Antioxidant,
Metabolic syndrome,
Robusta coffee leaves

DOI:

[https://doi.org/10.26656/fr.2017.4\(S3\).S10](https://doi.org/10.26656/fr.2017.4(S3).S10)

Abstract

Metabolic syndrome is a group of symptoms with obesity as the main determinant. Obesity increases oxidative stress which results in an imbalance between free radicals and antioxidants. Robusta coffee leaves are a source of antioxidants that can increase antioxidant levels in the body. The study was conducted to prove the effect of Robusta coffee leaves on the activity of superoxide dismutase (SOD) in metabolic syndrome rats. An experimental study of randomized pre-post test with control group design was conducted on 36 male Wistar rats. The rats besides the control group 1 were fed with high fat-fructose diet (HFFD) for 14 days. Samples were divided into 6 groups, each of 6 rats: control group 1 healthy rat without treatment (K1), control group 2 rats without treatment (K2), rats were given 20 mg/kg of mangiferin (X1), groups X2, X3, and X4 were given Robusta coffee leaves with each of dose is 0.09 g/200 g BW, 0.18 g/200 g BW and 0.36 g/200 gBW. Each dose of coffee leaves was steeped with 3.6 mL of water at 70°C for 10 mins. The intervention was given for 28 days. The metabolic syndrome was achieved with blood glucose level 131.14-132.70 mg/dL, triglycerides 153.49-157.18 mg/dL, and HDL 24.59-26.22 mg/dL. The highest increased in SOD levels in group X4 (35.78±8.27 u/mL) ($p=0.000$). The intervention of Robusta coffee leaves 0.36 g/200 g BW is the most effective dose in increasing SOD levels in metabolic syndrome rats.

1. Introduction

Metabolic syndrome is a cluster of conditions including obesity, dyslipidemia, hyperglycemia, and high blood pressure (Lam and LeRoith, 2019). Based on National Cholesterol Education Program which is modified for the Asian region define metabolic syndrome as having three or more of conditions, such as waist circumference of more than 90 cm in men and more than 80 cm in women; triglyceride level over 150 mg/dL; high-density lipoprotein (HDL) less than 45 mg/dL for men and less than 50 mg/dL for women; blood pressure of 130/85 mmHg or higher; and fasting glucose level more than 110 mg/dL (Kaur, 2014).

The prevalence of metabolic syndrome in the world is between 10% and 84% based on ethnicity, age, gender, and race. The prevalence of metabolic syndrome in

America in 2000 was about 47 million adult population. From 1988-1994 to 2007-2012, the prevalence of metabolic syndrome in America increased from 25.3% to 34.2% in adults aged 18 years or older (Moore *et al.*, 2017). In China, the incidence of metabolic syndrome increased by 37% during 2001-2009 (Guo *et al.*, 2017). The prevalence of metabolic syndrome in Indonesia increased by 23% in 2005 (Suhaema and Masthalina, 2015).

Obesity is a major determinant of metabolic syndrome. Obesity is associated with systemic inflammation in adipose tissue which increases pro-inflammatory status and oxidative stress (Marseglia *et al.*, 2014). Oxidative stress occurs because of an imbalance between free radicals and antioxidants in the body. Free radicals are atoms or molecules that have unpaired electrons. Free radicals are divided into two

*Corresponding author.

Email: widyastutinurmasari@gmail.com

categories, Reactive Oxygen Species (ROS) and Reactive Nitrogen Species (RNS). Reactive Oxygen Species (ROS) are reactive oxygen that is physiologically formed in cells as a by-product of cell metabolism. Oxidative damage is more prone to occur in obese people because of decreased sources of antioxidants in the body (Widayati, 2012; Marsegia et al., 2014).

Superoxide dismutase (SOD), catalase, and glutathione peroxidase (GPx) are endogenous antioxidants that act as ROS scavengers (Incalza et al., 2018). Superoxide dismutase (SOD) is the most powerful antioxidant in cells that acts as the main defense against free radicals (Ighodaro and Akinloye, 2018). When endogenous antioxidants are inadequate to eliminate ROS from the body, the body needs exogenous antioxidants, antioxidants derived from food (Mironczuk-Chodakowska et al., 2018). One of the food ingredients that contain antioxidants is coffee leaves (Huyut et al., 2017).

Coffee leaves have been consumed by people in Sumatra as a tea beverage, known as "Kawa Daun" or "Kopi Kawa" (Rasyid et al., 2013). Coffee leaves contain various phytochemicals, including alkaloids, flavonoids, terpenoids, amino acids, sucrose, tannins, xantonoids, phenolic acids, and catechins that function as anti-inflammatory, antidiabetic, antihyperlipidemic, antioxidant, antimicrobial, and neuroprotective which can control oxidative stress (Chen et al., 2018).

The most widely cultivated coffee plants in Indonesia is Robusta coffee (*Coffea canephora*) (Kanisius, 1988). Previous research describes the ethanolic extract of Robusta coffee leaves has an antioxidant activity with IC₅₀ value of 10.68 mg/mL. Ethanolic extract of Robusta coffee in mice with Type 2 Diabetes Mellitus by high fat and sucrose diet can lower blood glucose levels (Shiyan et al., 2017). Provision of 5% Robusta coffee leaves infusion can reduce blood glucose levels in alloxan-induced rats (Purwaningsih, 2014).

Brewed Robusta coffee leaves are known to become a traditional drink by people in several regions in Indonesia. The content in Robusta coffee leaves contains phytochemical compounds that make Robusta coffee leaves into food material that can be used as a source of antioxidants. The aim of this study was to examine the effect of brewed Robusta coffee leaves on SOD levels in metabolic syndrome rats.

2. Materials and methods

2.1 Materials

The Robusta coffee leaves were hand-picked from Mekarsari Village, Pasir Jambu District, Bandung Regency, West Java, Indonesia.

2.2 Methods

All experiments were approved by The Ethical Committee of Medical Research of Faculty of Medicine, Universitas Diponegoro (No. 16/EC/H/FK-UNDIP/III/2019). This study was a randomized pre-posttest with control group design. The study was conducted in March-May 2019 in Centre for Food and Nutrition of Gajah Mada University, Yogyakarta-Indonesia (Pusat Studi Pangan dan Gizi Universitas Gajah Mada, Yogyakarta-Indonesia) for the maintenance of rats and examination of SOD levels. The Tea and Quinine Research Center Gambung, Bandung-Indonesia (Pusat Penelitian Teh dan Kina Gambung, Bandung-Indonesia) for manufacturing brewed Robusta coffee leaves products.

The Robusta coffee leaves were processed as Japanese style Green Tea Process (JGTP). The leaves were blanched in boiling water at 100°C for 75 s, then dipped in water. Coffee leaves were drained and cooled, and the midrib removed. Furthermore, the coffee leaves were crushed three times using a Crushing Tearing and Curling (CTC) machine. The coffee leaves that have been crushed were dried using a rack drier at 80°C for 4-5 hrs. Coffee leaves were processed in a mini-processing green tea processing laboratory in The Tea Quality Processing and Testing Laboratory in The Tea and Quinine Research Center Gambung, Bandung-Indonesia (Atmaja et al., 2018).

Thirty-six male Wistar rats (6 weeks old and 150-200 g body weight) were randomly divided into 6 groups of 6 rats in each group after a week of adaptation. All rats were housed individually. The rats were all fed with a standard chow (Comfeed II) 20 g/day and drink ad libitum. The rats besides the control group 1 were fed with high fat-fructose diet (HFFD), contains pork oil (20%), cholesterol (1.5%), cholic acid (0.5%) and a standard chow (80%) given orally, while fructose of 1 mL/ 200gBW is given through sonde for 14 days.

Rats were conditioned into metabolic syndrome with blood glucose ≥ 110 mg/dL, triglycerides >150 mg/dL and HDL <40 mg/dL (Rochlani et al., 2017; Saifur Rohman et al., 2017). The control group 1 healthy rats without treatment (K1), control group 2 metabolic syndrome rats without treatment (K2), metabolic syndrome rats were given 20 mg/kg of mangiferin (X1), groups X2, X3, and X4 metabolic syndrome rats were given Robusta coffee leaves with each of dose is 0.09

g/200gBW, 0.18 g/200 gBW and 0.36 g/200 gBW. Each dose of coffee leaves was brewed with 3.6 mL of water at 70°C for 10 mins. The intervention was given for 28 days.

Measurement of blood glucose, triglycerides, and HDL as determinants of metabolic syndrome criteria in rats was carried out after 14 days of HFFD administration. Measurement of rat body weight was done once a week and measurements of SOD levels were carried out before and after the intervention. Examination of SOD levels uses the Enzyme-Linked Immunosorbent Assay (ELISA) method by taking blood serum in the retro-orbital plexus.

2.3 Data analysis

The results were expressed in mean \pm standard deviation (SD) and some in the median (minimum-maximum). The normality of the study was tested using the Kolmogorov-Smirnov test. Paired t-test and one-way analysis of variance (ANOVA) were used for parametric results; differences between the groups were evaluated by post-hoc test. Wilcoxon test, Kruskal Wallis, and Mann-Whitney were used for non-parametric results.

3. Results

3.1 Characteristics of subject

High fat-fructose diet administration for 14 days increased blood glucose ≥ 110 mg/dL, triglycerides >150 mg/dL and decrease HDL <40 mg/dL. The conditioning of rats into metabolic syndrome was achieved with blood glucose 131.14-132.70 mg/dL, triglyceride 153.59-157.18 mg/dL and 24.59-2.22 mg/dL as in Table 1.

Table 1. Characteristics of subject after high fat-fructose diet (HFFD) administration

Groups	Body Weight (g)	Blood Glucose (mg/dL)	Triglyceride (mg/dL)	HDL (mg/dL)
K1	180.50 \pm 3.06 ¹	71.29 \pm 1.53 ¹	68.77 \pm 5.97 ¹	86.36 \pm 2.28 ¹
K2	189.91 \pm 2.08 ¹	132.70 \pm 1.48 ¹	157.18 \pm 4.88 ¹	25.05 \pm 1.84 ¹
X1	191.25 \pm 5.59 ¹	131.81 \pm 1.88 ¹	153.75 \pm 3.11 ¹	26.22 \pm 1.69 ¹
X2	190.66 \pm 3.64 ¹	131.56 \pm 2.57 ¹	156.12 \pm 2.48 ¹	26.22 \pm 1.30 ¹
X3	192.50 (184-195.5) ²	131.14 \pm 2.13 ¹	153.49 \pm 1.96 ¹	26.22 \pm 2.15 ¹
X4	195.66 \pm 3.01 ¹	132.53 \pm 2.36 ¹	156.12 \pm 2.77 ¹	24.59 \pm 1.99 ¹

¹Mean \pm SD, ²Median (minimum-maximum)

Table 2. Characteristics of body weight before and after brewed Robusta coffee leaves treatment

Groups	Body Weight (g)			p
	Before Treatment	After Treatment	Delta	
K1	180.50 \pm 3.06 ^{1,a}	198.75 \pm 2.81 ^{1,a}	18.25 \pm 1.16 ^{1,a}	0
K2	189.91 \pm 2.08 ^{1,b}	224.12 \pm 2.11 ^{1,c}	34.20 \pm 1.08 ^{1,b}	0
X1	191.25 \pm 5.59 ^{1,b}	212.25 \pm 5.94 ^{1b}	20.12 (20-24) ^{2,c}	0
X2	190.66 \pm 3.64 ^{1,b}	218.83 \pm 3.62 ^{1,b,c}	28.16 \pm 0.71 ^{1,d}	0
X3	192.50 (184-195.5) ^{2,b}	217.33 \pm 4.66 ^{1,b,c}	25.58 \pm 2.35 ^{1,d,e}	0.028
X4	195.66 \pm 3.01 ^{1,b}	219.12 \pm 4.11 ^{1,b,c}	23.45 \pm 1.39 ^{1,c,e}	0
p	0.001*	0.000*	0.000*	

¹Mean \pm SD, ²Median (minimum-maximum) *Significant (p \leq 0.05) ^{a,b,c,d,e} Post Hoc Test

3.2 Characteristics of body weight before and after brewed Robusta coffee leaves treatment

Rats body weights were significantly increased in all groups (p \leq 0.000). There was a significant difference in body weight between groups before treatment and after treatment (p=0.000). X2 group increased body weight (28.16 \pm 0.71 g) higher than X3 (25.58 \pm 2.35 g) and X4 (23.45 \pm 1.39 g) groups as shown in Table 2.

3.3 Superoxide dismutase (SOD) levels before and after brewed Robusta coffee leaves treatment

There were significantly increased on SOD levels in mangiferin treatment group (X1) (p=0.000), 0.18 g/200 gBW Robusta coffee leaves treatment group (X3) (p=0.000), and 0.36 g/200 g BW Robusta coffee leaves treatment group (X4) (p=0.000). SOD levels between groups before treatment and after treatment were significantly different. X4 group with 0.36 g/200 g BW Robusta coffee leaves increased SOD levels (35.78 \pm 8.27 u/mL) higher than X2 (6.86 \pm 7.63 u/mL) and X3 (19.30 \pm 5.76 u/mL) groups. X1 group with mangiferin treatment had increased SOD levels (35.40 \pm 8.33 u/mL) almost the same as X4 group as seen in Table 3.

4. Discussion

This study showed that HFFD administration increase in body weight, blood glucose, triglycerides, and decreased HDL. Previous studies have explained that HFFD administration can induce weight gain, glucose intolerance, and leptin intolerance (Jarukumjorn et al., 2016). HFFD administration in rats for 14 days can result in hyperglycemia and dyslipidemia (Octavia et al., 2017).

Table 3. Superoxide Dismutase (SOD) before and after brewed Robusta coffee leaves treatment

Groups	SOD Levels (u/mL)			p
	Before Treatment	After Treatment	Delta	
K1	79.41±3.66 ^{1,a}	78.70±3.46 ^{1,a}	0.70±1.22 ^{1,a,b}	0.216
K2	25.49±6.32 ^{1,b}	19.44±4.49 ^{1,b}	-2.99 (-22.33-(-1.20)) ^{2,b}	0.128
X1	29.41±4.63 ^{1,b}	64.81±4.82 ^{1,c}	35.40±8.33 ^{1,c}	0.000
X2	26.47±3.66 ^{1,b}	33.33±5.23 ^{1,c}	6.86±7.63 ^{1,c}	0.079
X3	24.83±6.40 ^{1,b}	44.13±5.15 ^{1,d}	19.30±5.76 ^{1,d}	0.000
X4	22.55±6.41 ^{1,b}	58.33±3.46 ^{1,c}	35.78±8.27 ^{1,c}	0.000
p	0.000*	0.000*	0.000*	

¹Mean±SD, ²Median (minimum-maximum) *Significant (p<0.05) ^{a,b,c,d,e} Post Hoc Test

High-fructose diet results in the accumulation of triglycerides and cholesterol which causes a decrease in insulin sensitivity, insulin resistance, and glucose intolerance (Wong *et al.*, 2016). Previous studies have explained that fructose diet results in an increase in body weight in mice (Jürgens *et al.*, 2005). High-fat diet results in an increase in body mass, triglycerides, and LDL (Fraulob *et al.*, 2010).

Brewed Robusta coffee leaves were processed using the Japanese style Green Tea Process (JGTP) method. This process maintains the phytochemical content in coffee leaves. Previous studies have explained that the JGTP method in Arabica coffee leaves has a higher phytochemical concentration compared to other methods (Chen *et al.*, 2018).

After the intervention, body weight showed a significant increased between groups before and after treatment. In the K1 group, healthy rats were only given standard chow and drink experienced an increase in body weight. Weight gain in the treatment group was lower than in the K2 group. Rats with a dose of 0.36 g/200 g BW of Robusta coffee leaves experienced the lowest weight gain compared with a dose of 0.09 g/200 g BW and 0.18 g/200 g BW of Robusta coffee leaves. The higher the doses administered, the lower was the increase in body weight of rats. This relates to the antioxidant content in Robusta coffee leaves which can reduce weight gain. Antioxidants that may be present in Robusta coffee leaves are alkaloids, flavonoids, xantonoids, and phenolics (Chen *et al.*, 2018).

Based on previous research, the administration of flavonoids can maintain body weight and reduce the absorption of cholesterol in the gastrointestinal (Bertoia *et al.*, 2016). Other studies explain that the administration of alkaloids and isoflavones can reduce body weight, triglycerides, total cholesterol, LDL, and significantly increase HDL levels (Hurt and Wilson., 2012; Adeneye and Crooks., 2015). Chlorogenic acid which is included as phenolic acid can also reduce body weight in obese rats (Wang *et al.*, 2019).

Rats with mangiferin treatment increased body

weight lower than brewed Robusta coffee leaves. This is because mangiferin, which belongs to the xantonoid group, can reduce weight, total free fatty acids, and triglyceride levels. Weight loss mainly occurs in fat mass. Mangiferin regulates enzymes in carbohydrate oxidation in muscles and lipid utilization pathways in hepatocytes. Mangiferin also suppresses the lipogenesis enzyme in hepatocytes. This compound can prevent weight gain because of its ability to stimulate the utilization of carbohydrates and fats while inhibiting lipid synthesis (Ekaterina *et al.*, 2016). Mangiferin in this study acted as the standard antioxidant compound.

SOD levels between groups before and after administration of brewed Robusta coffee leaves increased. The administration of brewed Robusta coffee leaves at a dose of 0.36 g/200 g BW increased SOD levels in rats higher than the administration of brewed doses of 0.09 g/200 g BW and 0.18 g/200 g BW of Robusta coffee leaves. This shows that the higher the doses administered, the greater was the increase in the SOD level in rats.

The K1 group was only given a standard diet where the SOD levels in this group experienced an insignificant increase. K2 group was given a standard diet without treatment. SOD levels in this group decreased. Metabolic syndrome results in decreased antioxidant activity in the body, such as SOD and GPx. This is related to the high oxidative stress in sufferers of metabolic syndrome (Baez-Duarte *et al.*, 2016). Cells can maintain a state of reduction-oxidation (redox) when the production of ROS in cells is low or moderate if there is an increase in ROS it will burden antioxidant defenses causing oxidative stress and cell damage. The disruption of redox conditions causes poisoning of cells through the production of peroxidase and free radicals (Bresciani *et al.*, 2015).

Group X1 was given a standard diet and mangiferin. SOD levels in this group experienced a significant increase. Mangiferin is thought to be found in Robusta coffee leaves as found in Arabica coffee leaves content. This compound belongs to the xantonoid group and has a

strong antioxidant activity because it can scavenge ROS by initiating lipid peroxidation. Mangiferin can neutralize some reactive species and influence the expression and activity of the detoxification enzyme (Fomenko and Chi, 2016). This can reduce oxidative stress and inflammation. These compounds maintain the balance of antioxidant enzymes, such as SOD, GPx, and catalase and can prevent the production of hydroxyl radicals due to the ability of chelating iron in the Fenton reaction. The Fenton reaction involves the production of hydroxyl radicals and the oxidation of Fe^{2+} to Fe^{3+} . Mangiferin inhibits the Fenton reaction by correlating Fe^{2+} ions thereby reducing ROS production (Arozal et al., 2015; Gold-Smith et al., 2016).

Group X2, X3 and X4 showed increases in SOD levels. The increase in SOD levels in the three groups was thought to be due to the antioxidant content found in Robusta coffee leaves such as the content contained in Arabica coffee leaves. Robusta coffee leaves contain phenolic content of 27.04 $\mu\text{g/g}$ and flavonoids of 10.90 $\mu\text{g/g}$. Antioxidant activity in Robusta coffee leaves extract showed inhibition of 79.43% (Nayeem et al., 2011). Whereas, Arabica coffee leaves contained caffeine content of 7.14 mg/g, chlorogenic acid 21.39 mg/g, and mangiferin 7.88 mg/g coffee leaves processed by the JGTP method (Chen et al., 2018).

The antioxidant content in Robusta coffee leaves, in addition to mangiferin, is phenolic. This compound can inhibit oxidation, scavenge free radicals, and act as chelating agents which can stop progressive auto-oxidative damage. In addition to phenolic, there is flavonoids that can increase SOD levels in rats (Brewer, 2011). This is because flavonoids work as antioxidants that protect the body against ROS. Flavonoids directly scavenge free radicals and act as intracellular antioxidants which can inhibit free radical producing enzymes. Flavonoids stabilize free radicals by reacting to the reactive component of free radicals. High reactivity in the hydroxyl flavonoid group makes the radicals inactive (Banjarnahor and Artanti., 2014; Panche et al., 2016). Research conducted on rats by giving flavonoids with a dose of 100 mg/kg can increase SOD levels (Luo et al., 2014).

Another antioxidant in coffee leaves is caffeine. Caffeine is a derivative of xanthine which has a role to directly inhibit lipid peroxidation and a high level of inhibition of radical formation (Tellone et al., 2015). This compound can reduce oxidative stress and ROS and protect the antioxidant system (Jeszka-Skowron et al., 2016). Caffeine captures hydroxyl radicals and superoxide by measuring O_2^- after reacting with caffeine which is measured using electron paramagnetic

resonance (Yamagata, 2018).

Other antioxidant compounds in Robusta coffee leaves are chlorogenic acid which acts as an antioxidant by donating hydrogen atoms to reduce free radicals and inhibit oxidation reactions. After donating a hydrogen atom, chlorogenic acid will be oxidized to phenoxyl radicals and stabilized by resonance stabilization (Liang and Kitts, 2016). This compound scavenges ROS resulting from the consumption of a high-fat diet by suppressing the expression of inflammation through inhibition of the protein tyrosine phosphatase 1B, resulting in reduced fat accumulation, weight loss, and reduced risk of insulin resistance (Buscemi et al., 2016; Santana-Gálvez et al., 2017).

5. Conclusion

Robusta coffee leaves processed by the Japanese style green tea process contain antioxidants that are useful in the treatment of metabolic syndrome. The administration of brewed Robusta coffee leaves significantly increase SOD levels in metabolic syndrome rats with the most effective dose is 0.36 g/200 g BW.

Conflict of Interest

The authors declare no conflict of interest.

Acknowledgments

This research was supported by Research and Development (RPP) from the Faculty of Medicine, Universitas Diponegoro.

References

- Adeneye, A.A. and Crooks, P.A. (2015). Weight losing, antihyperlipidemic and cardioprotective effects of the alkaloid fraction of *Hunteria umbellata* seed extract on normal and triton-induced hyperlipidemic rats. *Asian Pacific Journal of Tropical Biomedicine*, 5(5), 387–394. [https://doi.org/10.1016/S2221-1691\(15\)30374-9](https://doi.org/10.1016/S2221-1691(15)30374-9)
- Arozal, W., Suyatna, F., Juniantito, V., Rosdiana, D., Amurugam, S., Aulia, R., Monayo, E. and Siswandi, R. (2014). The Effects of Mangiferin (*Mangifera indica* L) in Doxorubicin-induced Cardiotoxicity in Rats. *Drug Research*, 65(11), 574–580. <https://doi.org/10.1055/s-0034-1394457>
- Atmaja, M.I.P., Shabri, Harianto, S., Maulana, H. and Rohdiana, D. (2018). Physical Characteristics of Green Tea Powder Processed by Disc and Stone Mills. *Jurnal Teknologi dan Industri Pangan*, 29, 77–84. <https://doi.org/10.6066/jtip.2018.29.1.77>
- Baez-Duarte, B.G., Zamora-Ginez, I., De Jesús, K.L.,

- Torres-Rasgado, E., González-Mejía, M.E., Porchia, L., Ruiz-Vivanco, G. and Pérez-Fuentes, R. (2016). Association of the Metabolic Syndrome with Antioxidant Defense and Outstanding Superoxide Dismutase Activity in Mexican Subjects. *Metabolic Syndrome and Related Disorders*, 14(3), 154–160. <https://doi.org/10.1089/met.2015.0088>
- Banjarnahor, S.D.S. and Artanti, N. (2015). Antioxidant properties of flavonoids. *Medical Journal of Indonesia*, 23(4), 239–244. <https://doi.org/10.13181/mji.v23i4.1015>
- Bertoia, M.L., Rimm, E.B., Mukamal, K.J., Hu, F.B., Willett, W.C. and Cassidy, A. (2016). Dietary flavonoid intake and weight maintenance: three prospective cohorts of 124 086 US men and women followed for up to 24 years. *BMJ*, 2016(352), i17. <https://doi.org/10.1136/bmj.i17>
- Bresciani, G., da Cruz, I.B.M. and González-Gallego, J. (2015). Manganese Superoxide Dismutase and Oxidative Stress Modulation. In Makowski, G.S. (Ed.) *Advances in Clinical Chemistry*, p. 87–130. USA: Elsevier. <https://doi.org/10.1016/bs.acc.2014.11.001>
- Brewer, M.S. (2011). Natural Antioxidants: Sources, Compounds, Mechanisms of Action, and Potential Applications. *Comprehensive Reviews in Food Science and Food Safety*, 10(4), 221–247. <https://doi.org/10.1111/j.1541-4337.2011.00156.x>
- Buscemi, S., Marventano, S., Antoci, M., Cagnetti, A., Castorina, G., Galvano, F., Marranzano, M. and Mistretta, A. (2016). Coffee and metabolic impairment: An updated review of epidemiological studies. *NFS Journal*, 3, 1–7. <https://doi.org/10.1016/j.nfs.2016.02.001>
- Chen, X.-M., Ma, Z. and Kitts, D.D. (2018). Effects of processing method and age of leaves on phytochemical profiles and bioactivity of coffee leaves. *Food Chemistry*, 249, 143–153. <https://doi.org/10.1016/j.foodchem.2017.12.073>
- Fomenko, E.V. and Chi, Y. (2016). Mangiferin modulation of metabolism and metabolic syndrome. *BioFactors*, 42(5), 492–503. <https://doi.org/10.1002/biof.1309>
- Fraulob, J.C., Ogg-Diamantino, R., Fernandes-Santos, C., Aguila, M.B. and Mandarim-de-Lacerda, C.A. (2010). A Mouse Model of Metabolic Syndrome: Insulin Resistance, Fatty Liver and Non-Alcoholic Fatty Pancreas Disease (NAFPD) in C57BL/6 Mice Fed a High Fat Diet. *Journal of Clinical Biochemistry and Nutrition*, 46(3), 212–223. <https://doi.org/10.3164/jcbrn.09-83>
- Gold-Smith, F., Fernandez, A. and Bishop, K. (2016). Mangiferin and Cancer: Mechanisms of Action. *Nutrients*, 8(7), 396. <https://doi.org/10.3390/nu8070396>
- Guo, H., Gao, X., Ma, R., Liu, J., Ding, Y., Zhang, M., Zhang, J., Mu, L., He, J., Yan, Y., Ma, J., Guo, S. and Wei, S. (2017). Prevalence of Metabolic Syndrome and its Associated Factors among Multi-ethnic Adults in Rural Areas in Xinjiang, China. *Scientific Reports*, 7(1), 17643. <https://doi.org/10.1038/s41598-017-17870-5>
- Hurt, R.T. and Wilson, T. (2012). Geriatric Obesity: Evaluating the Evidence for the Use of Flavonoids to Promote Weight Loss. *Journal of Nutrition in Gerontology and Geriatrics*, 31(3), 269–289. <https://doi.org/10.1080/21551197.2012.698222>
- Huyut, Z., Beydemir, Ş. and Gülçin, İ. (2017). Antioxidant and Antiradical Properties of Selected Flavonoids and Phenolic Compounds. *Biochemistry Research International*, 2017, 1–10. <https://doi.org/10.1155/2017/7616791>
- Ighodaro, O.M. and Akinloye, O.A. (2018). First line defence antioxidants-superoxide dismutase (SOD), catalase (CAT) and glutathione peroxidase (GPX): Their fundamental role in the entire antioxidant defence grid. *Alexandria Journal of Medicine*, 54(4), 287–293. <https://doi.org/10.1016/j.ajme.2017.09.001>
- Imran, M., Arshad, M.S., Butt, M.S., Kwon, J.-H., Arshad, M.U. and Sultan, M.T. (2017). Mangiferin: a natural miracle bioactive compound against lifestyle related disorders. *Lipids in Health and Disease*, 16(1), 84. <https://doi.org/10.1186/s12944-017-0449-y>
- Incalza, M.A., D'Oria, R., Natalicchio, A., Perrini, S., Laviola, L. and Giorgino, F. (2018). Oxidative stress and reactive oxygen species in endothelial dysfunction associated with cardiovascular and metabolic diseases. *Vascular Pharmacology*, 100, 1–19. <https://doi.org/10.1016/j.vph.2017.05.005>
- Jarukamjorn, K., Jearapong, N., Pimson, C. and Chatuphonprasert, W. (2016). A High-Fat, High-Fructose Diet Induces Antioxidant Imbalance and Increases the Risk and Progression of Nonalcoholic Fatty Liver Disease in Mice. *Scientifica*, 2016, 1–10. <https://doi.org/10.1155/2016/5029414>
- Jeszka-Skowron, M., Sentkowska, A., Pyrzyńska, K. and De Peña, M.P. (2016). Chlorogenic acids, caffeine content and antioxidant properties of green coffee extracts: influence of green coffee bean preparation. *European Food Research and Technology*, 242(8), 1403–1409. <https://doi.org/10.1007/s00217-016-2643-y>
- Jürgens, H., Haass, W., Castañeda, T.R., Schürmann, A.,

- Koebnick, C., Dombrowski, F., Otto, B., Nawrocki, A., Scherer, P., Spranger, J., Ristow, M., Joost, H., Havel, P. and Tschöp, M.H. (2005). Consuming Fructose-sweetened Beverages Increases Body Adiposity in Mice. *Obesity Research*, 13(7), 1146–1156. <https://doi.org/10.1038/oby.2005.136>
- Kanisius, A. A. (1988). *Budidaya Tanaman Kopi*. 16th ed. Yogyakarta: Kanisius.
- Kaur, J. (2014). A Comprehensive Review on Metabolic Syndrome. *Cardiology Research and Practice*, 2014, 1–21. <https://doi.org/10.1155/2014/943162>
- Lam, D.W. and LeRoith, D. (2019). Metabolic Syndrome. In *Endotext.*, p. 1–41. Retrieved from <https://www.ncbi.nlm.nih.gov/books/NBK278936/>
- Liang, N. and Kitts, D.D. (2016). Role of Chlorogenic Acids in Controlling Oxidative and Inflammatory Stress Conditions. *Nutrients*, 8(16), 1–20.
- Luo, A., Peng, P., Fei, W., Yang, L. and Fan, Y. (2014). Isolation and Antioxidant Activity (In Vitro and In Vivo) of The Flavonoid From Tartarian-buckwheat. *Journal of Scientific and Innovative Research*, 3(2), 168–172.
- Marseglia, L., Manti, S., D'Angelo, G., Nicotera, A., Parisi, E., Di Rosa, G., Gitto, E. and Arrigo, T. (2014). Oxidative Stress in Obesity: A Critical Component in Human Diseases. *International Journal of Molecular Sciences*, 16(1), 378–400. <https://doi.org/10.3390/ijms16010378>
- Mironczuk-Chodakowska, I., Witkowska, A.M. and Zujko, M.E. (2018). Endogenous non-enzymatic antioxidants in the human body. *Advances in Medical Sciences*, 63(1), 68–78. <https://doi.org/10.1016/j.advms.2017.05.005>
- Moore, J.X., Chaudhary, N. and Akinyemiju, T. (2017). Metabolic Syndrome Prevalence by Race/Ethnicity and Sex in the United States, National Health and Nutrition Examination Survey, 1988–2012. *Preventing Chronic Disease*, 14(24), 160287. <https://doi.org/10.5888/pcd14.160287>
- Nayeem, N., Denny, G. and Mehta, S.K. (2011). Comparative phytochemical analysis, antimicrobial and anti oxidant activity of the methanolic extracts of the leaves of Coffea Arabica and Coffea Robusta. *Der Pharmacia Lettre*, 3(1), 292–297.
- Octavia, Z.F., Djamiatun, K. and Suci, N. (2017). The effect of synbiotic yogurt of tanduk banana fl our in lipid profile of metabolic syndrome rats. *Jurnal Gizi Klinik Indonesia*, 13(4), 159–169. <https://doi.org/10.22146/ijcn.19369>
- Panche, A.N., Diwan, A.D. and Chandra, S.R. (2016). Flavonoids: an overview. *Journal of Nutritional Science*, 5(47), e47. <https://doi.org/10.1017/jns.2016.41>
- Purwaningsih, S. (2014). Effect of Infusion Coffee Leaves (*Coffea robusta* Lind.) on Blood Glucose Level on the Mice in Alloxan Induced. *Veterinaria Medika*, 7(3), 208–215.
- Rasyid, R., Sanjaya, W.F. and Zulharmita. (2013). Penetapan Kadar Kofein Daun Kopi Kawa (*Coffea Robusta*, Lind). *Jurnal Farmasi Higea*, 5(2), 137–143. [In Bahasa Indonesia].
- Rochlani, Y., Pothineni, N.V., Kovelamudi, S. and Mehta, J.L. (2017). Metabolic syndrome: pathophysiology, management, and modulation by natural compounds. *Therapeutic Advances in Cardiovascular Disease*, 11(8), 215–225. <https://doi.org/10.1177/1753944717711379>
- Saifur Rohman, M., Lukitasari, M., Adi Nugroho, D., Nashi, W., Ida Panca Nugraheini, N. and Wahyu Sardjono, E. (2017). Development of an Experimental Model of Metabolic Syndrome in Sprague Dawley Rat. *Research Journal of Life Science*, 4(1), 76–86. <https://doi.org/10.21776/ub.rjls.2017.004.01.10>
- Santana-Gálvez, J., Cisneros-Zevallos, L. and Jacobo-Velázquez, D. (2017). Chlorogenic Acid: Recent Advances on Its Dual Role as a Food Additive and a Nutraceutical against Metabolic Syndrome. *Molecules*, 22(3), 358. <https://doi.org/10.3390/molecules22030358>
- Shiyan, S., Herlina, Arsela, D. and Latifah, E. (2017). Aktivitas Antidiabetes Ekstrak Etanolik Daun Kopi Robusta (*Coffea canephora*) Pada Tikus Diabeets Tipe 2 Yang Diberi Diet Lemak Tinggi dan Sukrosa. *Jurnal Farmasi Sains Dan Praktis*, III(2), 39–46. [In Bahasa Indonesia].
- Suhaema. and Masthalina, H. (2015). Pola Konsumsi dengan Terjadinya Sindrom Metabolik di Indonesia (Consumption Patterns with Occurrence of Metabolic Syndrome in Indonesia). *Jurnal Kesehatan Masyarakat Nasional*, 9(4), 340–347. [In Bahasa Indonesia].
- Tellone, E., Galtieri, A., Giardina, B., Russo, A., Bellocco, E., Barreca, D. and Ficarra, S. (2015). Antioxidant Activity of Caffeine. In Preedy, V.R. (Ed.) *Coffee in Health and Disease Prevention.*, p. 835–842. USA: Academic Press. <https://doi.org/10.1016/B978-0-12-409517-5.00092-9>
- Wang, Z., Lam, K., Hu, J., Ge, S., Zhou, A., Zheng, B., Zeng, S. and Lin, S. (2019). Chlorogenic acid alleviates obesity and modulates gut microbiota in high-fat-fed mice. *Food Science and Nutrition*, 7(2), 579–588. <https://doi.org/10.1002/fsn.3.868>
- Widayati, E. (2012). *Oxidasi Biologi, Radikal Bebas,*

- dan Antioxidant (Biological Oxidation, Free Radicals, and Antioxidants). *Majalah Islam Sultan Agung*, 50(128), 26–32.
- Wong, S.K., Chin, K.-Y., Suhaime, F.H., Fairus, A. and Ima-Nirwana, S. (2016). Animal models of metabolic syndrome: a review. *Nutrition and Metabolism*, 13(1), 65. <https://doi.org/10.1186/s12986-016-0123-9>
- Yamagata, K. (2018). Do Coffee Polyphenols Have a Preventive Action on Metabolic Syndrome Associated Endothelial Dysfunctions? An Assessment of the Current Evidence. *Antioxidants*, 7 (2), 26. <https://doi.org/10.3390/antiox7020026>

The increasing of superoxide dismutase (SOD) in metabolic syndrome rats by robusta coffee leaves (Coffea canephora)

ORIGINALITY REPORT

22%

SIMILARITY INDEX

16%

INTERNET SOURCES

16%

PUBLICATIONS

5%

STUDENT PAPERS

PRIMARY SOURCES

1	ijpronline.com Internet Source	1 %
2	www.ncbi.nlm.nih.gov Internet Source	1 %
3	www.freshpatents.com Internet Source	1 %
4	Dorota Woźniak, Izabela Nawrot-Hadzik, Weronika Kozłowska, Sylwester Ślusarczyk, Adam Matkowski. "Chapter 23-1 Caffeoylquinic Acids", Springer Science and Business Media LLC, 2020 Publication	1 %
5	Siddhi Patil, M. Vedashree, Pushpa S. Murthy. "Valorization of coffee leaves as a potential agri-food resource: bio-active compounds, applications and future prospective", Planta, 2022 Publication	1 %
6	Submitted to University of Bridgeport Student Paper	1 %

7	"Pathophysiology of Obesity-Induced Health Complications", Springer Science and Business Media LLC, 2020 Publication	1 %
8	ddfv.ufv.es Internet Source	1 %
9	informaticsjournals.com Internet Source	1 %
10	Rahmah Rahmah, Yoni Astuti, Harsono Salimo, Eti Poncorini Pamungkasari, Brian Wasita. "Beneficial Effect of Mesona palustris BL: A Review on Human and Animal Intervention", Open Access Macedonian Journal of Medical Sciences, 2022 Publication	1 %
11	Inggita Kusumastuty, Dian Handayani, Shafira Hanifa, Melinda Lisan, Etik Sulistyowati. "The Effect of Brown Rice on Superoxide Dismutase Level and Non-alcoholic Fatty Liver in an Sprague–Dawley Rat Model of High-fat High-fructose Diet-induced Obesity", Open Access Macedonian Journal of Medical Sciences, 2020 Publication	<1 %
12	vbn.aau.dk Internet Source	<1 %

13	"Bioactive Molecules in Food", Springer Science and Business Media LLC, 2019 Publication	<1 %
----	---	------

14	docplayer.net Internet Source	<1 %
----	----------------------------------	------

15	Praphul Joshi, Kaushik Ghosh, Vivek S. Natarajan, Purnendu Mandal. "Impact of healthcare access and socio-economic conditions on prevalence of metabolic syndrome: evidence from 2015 BRFSS", International Journal of Business Excellence, 2020 Publication	<1 %
----	---	------

16	elearning.medistra.ac.id Internet Source	<1 %
----	---	------

17	paper.researchbib.com Internet Source	<1 %
----	--	------

18	Maria Angela Incalza, Rossella D'Oria, Annalisa Natalicchio, Sebastio Perrini, Luigi Laviola, Francesco Giorgino. "Oxidative stress and reactive oxygen species in endothelial dysfunction associated with cardiovascular and metabolic diseases", Vascular Pharmacology, 2017 Publication	<1 %
----	---	------

19	dergipark.org.tr Internet Source	<1 %
----	-------------------------------------	------

20	www.dailymail.co.uk Internet Source	<1 %
21	Aloysius A. Pramudita, Yuyu Wahyu, Syamsul Rizal, Murman D. Prasetio, Agung N. Jati, Restu Wulansari, Harfan H. Ryanu. "Soil Water Content Estimation With the Presence of Vegetation Using Ultra Wideband Radar-Drone", IEEE Access, 2022 Publication	<1 %
22	Dayi Ji, Qiang Wang, Tingting Lu, Haile Ma, Xiumin Chen. "The effects of ultrasonication on the phytochemicals, antioxidant, and polyphenol oxidase and peroxidase activities in coffee leaves", Food Chemistry, 2022 Publication	<1 %
23	Submitted to Liverpool John Moores University Student Paper	<1 %
24	bdtd.biblioteca.ufpb.br Internet Source	<1 %
25	lipidworld.biomedcentral.com Internet Source	<1 %
26	www.e-sciencecentral.org Internet Source	<1 %
27	www.hindawi.com Internet Source	<1 %

28

www.omicsonline.org

Internet Source

<1 %

29

Suhuan Mei, Xiumin Chen. "Investigation into the anti-inflammatory mechanism of coffee leaf extract in LPS-induced Caco-2/U937 co-culture model through cytokines and NMR-based untargeted metabolomics analyses", Food Chemistry, 2023

Publication

<1 %

30

ejournal.almaata.ac.id

Internet Source

<1 %

31

eprints.poltekkesjogja.ac.id

Internet Source

<1 %

32

www.jidmr.com

Internet Source

<1 %

33

etd.aau.edu.et

Internet Source

<1 %

34

svuijm.journals.ekb.eg

Internet Source

<1 %

35

www.frontiersin.org

Internet Source

<1 %

36

www.rjpbcs.com

Internet Source

<1 %

37

"Metal, Metal Oxides and Metal Sulphides for Biomedical Applications", Springer Science

<1 %

38

Rinta Amalia, Adriyan Pramono, Diana Nur Afifah, Etika Ratna Noer, Muflihatul Muniroh, Andri Cahyo Kumoro. "Mangrove fruit (*Bruguiera gymnorhiza*) increases circulating GLP-1 and PYY, modulates lipid profiles, and reduces systemic inflammation by improving SCFA levels in obese wistar rats", *Heliyon*, 2022

Publication

<1 %

39

Xiaolong Wang, Qing Song, Zhiyong Wang, Fang Han. "A novel extracellular copper/zinc superoxide dismutase identified from *Nibea albiflora* and its characteristics under ammonia/nitrite stress", *International Journal of Biological Macromolecules*, 2018

Publication

<1 %

40

[id-press.eu](https://www.idpress.eu)

Internet Source

<1 %

41

jtos.polban.ac.id

Internet Source

<1 %

42

posnutricao.ufv.br

Internet Source

<1 %

43

www2.mdpi.com

Internet Source

<1 %

44

"Technical Report on the notification of infusion from coffee leaves (*Coffea arabica* L. and/or *Coffea canephora* Pierre ex A. Froehner) as a traditional food from a third country pursuant to Article 14 of Regulation (EU) 2015/2283", EFSA Supporting Publications, 2020

Publication

<1 %

45

Mardi M. Algandaby. "Crocic prevents metabolic syndrome in rats via enhancing PPAR-gamma and AMPK", Saudi Journal of Biological Sciences, 2020

Publication

<1 %

46

Shimaa Mustafa Elshazly, Mahmoud Mohamed Abdelaal, Seba Hassan Attia, Eman Soliman. "Reno-protective effect of mangiferin against methotrexate-induced kidney damage in male rats: PPAR γ -mediated antioxidant activity", Saudi Pharmaceutical Journal, 2022

Publication

<1 %

47

Xiumin Chen, Kaiwen Mu, David D. Kitts. "Characterization of phytochemical mixtures with inflammatory modulation potential from coffee leaves processed by green and black tea processing methods", Food Chemistry, 2019

Publication

<1 %

48	Yan - Yan Hor, Cheong - Hwa Ooi, Lee - Ching Lew, Mohamad Hafis Jaafar et al. "The molecular mechanisms of probiotic strains in improving aging bone and muscle of D - galactose - induced aging rats", Journal of Applied Microbiology, 2020 Publication	<1 %
49	adoc.pub Internet Source	<1 %
50	dokumen.pub Internet Source	<1 %
51	febs.onlinelibrary.wiley.com Internet Source	<1 %
52	mail.scialert.net Internet Source	<1 %
53	rep.bioscientifica.com Internet Source	<1 %
54	usir.salford.ac.uk Internet Source	<1 %
55	vital.seals.ac.za:8080 Internet Source	<1 %
56	www.scitepress.org Internet Source	<1 %
57	www.sysrevpharm.org Internet Source	<1 %

58

"Nutritional Antioxidant Therapies: Treatments and Perspectives", Springer Science and Business Media LLC, 2017

Publication

<1 %

59

Azizah Hikma Safitri, Nurina Tyagita, Taufiqurrachman Nasihun. "Porang Glucomannan Supplementation Improves Lipid Profile in Metabolic Syndrome Induced Rats", JOURNAL OF NATURAL REMEDIES, 2017

Publication

<1 %

60

D R Fajar, Akrom, E Darmawan. "The influence of black cumin seed oil therapy with dosage of 1.5 mL/day and 3 mL/day to interleukin-21 (IL-21) expression of the patients with metabolic syndrome risk", IOP Conference Series: Materials Science and Engineering, 2017

Publication

<1 %

61

Submitted to Gyeongsang National University

Student Paper

<1 %

Exclude quotes On

Exclude matches Off

Exclude bibliography On

The increasing of superoxide dismutase (SOD) in metabolic syndrome rats by robusta coffee leaves (Coffea canephora)

GRADEMARK REPORT

FINAL GRADE

/0

GENERAL COMMENTS

Instructor

PAGE 1

PAGE 2

PAGE 3

PAGE 4

PAGE 5

PAGE 6

PAGE 7

PAGE 8