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MiRNA encoded PTEN's Impact on Clinical-Pathological Features and Prognosis in Osteosarcoma: a Systematic Review and Meta-Analysis

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Abstract

Background: Osteosarcoma (OSC) is considered one of the most common malignant bone tumours in adolescents. Due to OSC's poor prognosis, a comprehensive approach to exploring these aspects is highly needed to improve the survival probability of OSC. In this study, we tried to explore the significance of miRNA-encoded PTEN for clinical-pathological features and prognostic value in OSC. **Method:** We performed this systematic review and meta-analysis using articles and sources published between 2013 and 2023 from six databases

(Scopus, PubMed, ProQuest, Science Direct, Sage Pub, and Cochrane). Included studies were clinical crosssectional studies. Other study designs, articles not written in English, without full text, and not relevant—were excluded. Then, ROBINS-I is used to evaluate the distance. The results are constructed according to the PICOS criteria in a table. The expression of miRNA related to OSC is assessed in the meta-analysis as the main outcome to determine its ability as a diagnostic and prognostic agent for OSC. This systematic review followed the PRISMA guidelines. **Results:** A total of 17 studies were included in the final screening. The meta-analysis showed significantly increased (p < 0.00001) miRNA expression in patients with OSC compared to healthy controlled with pooled md (2.85) (95% CI: 2.69, 3.02; $I^2=22\%$, p=0.20), the high inverse correlation (p < 0.001) between miRNA and PTEN expression was shown as mean effect size (-0.681) (95% CI: -0.787, -0.536; $I^2 =$ 75%, p < 0.0001), and the prognostic evaluation of OS was significantly increased in low expression miRNA (p <0.00001) with pooled OR. **Conclusion:** Fiveteen miRNAs from 17 studies were found, and together with PTEN expression, they may serve as potential prognostic biomarkers for OSC. High-level levels of miRNA expression are correlated with low PTEN expression, leading to a bad prognosis for OSC.

1. Introduction

Osteosarcoma (OSC) is the most frequent bone malignancy in both children and adults. It is a malignant mesenchymal tumour that accounts for 20–40% of all occurrences of bone malignancies. The metaphysis of the long bones in the lower extremities is the most frequent site of OSC, which is most common in people between the ages of five and twenty-five [1]. The inferior extremities, particularly the distal femur, proximal tibia, and proximal humerus, are shown to be the site of 74.5% of cases. In the meantime, factors like as age, gender, tumour location, biomarker levels, onset, and metastatic presence all affect the prognosis of OS. $61\pm15\%$ is the 5-year relative survival rate for all age groups, varying based on disease onset and gender. However, this figure drastically decreases to $20\pm5\%$ in patients with metastasis. Approximately 10-20% of OSC patients experience metastasis, with the lungs being a common site of involvement [2].

The high fatality rate drives the need for a more comprehensive multidisciplinary approach to the diagnosis and

management of OSC. To date, several treatment options for OSC include surgical excision, radiotherapy, and multi-agent systemic therapy. Several evaluations are required to ascertain which therapy will yield the best response in the patients. Furthermore, invasive biopsy is still required for OSC diagnosis evaluation and confirmation [3,4]. However, due to variations in sample collection, the accuracy of the biopsy-derived diagnosis and prognosis can fluctuate. Therefore, the diagnostic approach is an important aspect to develop to achieve better prognosis, and one method with great potential is identifying biomarkers [5–7].

MicroRNAs (miRNAs) play crucial roles in gene regulation, including the modulation of tumor suppressor genes such as PTEN (Phosphatase and Tensin Homolog). PTEN is pivotal in controlling cell proliferation, survival, and migration, often disrupted in various cancers [8]. In the cancer microenvironment, miRNAs that target PTEN can significantly influence cancer progression and metastasis. The tumor microenvironment, characterized by inflammatory cytokines such as IL-1 (Interleukin-1) and IL-6 (Interleukin-6), can modulate miRNA expression, thereby affecting PTEN levels. IL-1 and IL-6 are upregulated in many cancers, contributing to a pro-inflammatory state that fosters tumor growth and immune evasion [9]. IL-1 can induce miRNAs that suppress PTEN, enhancing oncogenic pathways and tumor aggressiveness. Similarly, IL-6 signaling can activate STAT3, which in turn can regulate miRNAs targeting PTEN, further diminishing its tumor-suppressive effects. The interplay between these cytokines and miRNA-mediated PTEN regulation underscores a complex network where the inflammatory milieu can promote oncogenesis by modulating critical gene expressions [10]. Understanding this dynamic offers potential therapeutic insights, where targeting specific miRNAs or cytokine pathways could restore PTEN function and inhibit cancer progression [11].

Biomarkers themselves can be used as therapeutic, prognostic, and diagnostic agents. MicroRNAs (miRNAs) are a class of endogenous non-coding RNAs with lengths of 19-25 nt. They are involved in differentiation, cell death, and the cell cycle by regulating gene expression through direct inhibition and degradation of mRNA translation [12]. In malignancies, this inhibitory function makes them tumor suppressors and biomarkers for the diagnosis and prognosis of patients. PTEN is one of the major tumor suppressors that has been extensively studied for its effects on cancer development, such as breast cancer, prostate cancer, and lung cancer. PTEN (phosphatase and tensin homolog deleted on chromosome ten) is a candidate tumor suppressor located on chromosome 10q23. PTEN works in the PI3K/AKT regulatory pathway, a lipid kinase family axis with the PIP3 product, which is involved in controlling cell proliferation to prevent cell development into cancer. The increase and decrease in PTEN expression are regulated by a number of proteins in posttranslational, posttranscriptional, and transcriptional mechanisms. One of these proteins is miRNA, which suppresses PTEN expression posttranscriptionally. In this systematic review and meta-analysis, we summarize current knowledge on the correlation between miRNA-PTEN and its impact on the clinicopathological and prognostic aspects of OSC, which has not been previously explored within the scope of a wide range of articles.

2. Methods

Eligibility criteria

The Preferred Reporting Items for Systematic Reviews and Meta-Analyses (PRISMA) were used for this systematic review [13]. From 2013 to 2023 (last search date September 20th, 2023), we have included original content. The study included original research publications that met the inclusion criteria for autologous clinical cross-sectional. Technical reports, editor's responses, narrative reviews, systematic reviews, meta-analyses, non-comparative studies, in silico studies, in vitro studies, in vivo studies, applied Scientific posters, research proposals, and conference abstracts were all diminished. Articles not written in English, with incomplete content, and not related to the miRNA and PTEN Gene expression in correlation to OSC as well as survival rate were also eliminated. The desired PICO criteria of the selected products included i) patients with OSC; ii) miRNA and PTEN expression evaluation towards OSC patients and its survival rate; iii) Comparison, normal cells either obtain from the same patients or other normal patients; and iv) Results, miRNA and PTEN evaluation utilizing RT-qPCR and two years survival rate.

Data search and selection

Research for this study were gathered through Scopus, PubMed, ProQuest, Science Direct, Sage Pub, Cochrane database searches. We used the following combined keywords to capture all potentially eligible literature in each databases: "(osteosarcoma) AND (miRNA OR microRNA) AND (PTEN) AND (clinicopathology OR prognosis)". The database was searched from its establishment until September 20th 2023 during 10-year periods prior to this review. The boolean operator was utilized among the Medical Subject Headings (MeSH) keywords determined from national institute of health (NIH) national library of medicine browser. The studies were kept using Mendeley Group Reference Manager in the authors' library. Clinical type articles were used as a filter in the Pubmed database, the the research article type was used for the Scopus, PubMed, ProQuest, Science Direct, Sage Pub, Cochrane database filter. All of these processes for article selection were conducted by the two independent authors (DRPR, KCT) who performed the literature searching process. The identified articles from those databases were first filtered according to their titles and abstracts, and then duplicates were eliminated. The same two authors conducted a second round of full-text evaluations for these publications that made it past the first screening round to determine whether they met our inclusion/exclusion criteria. All differences were addressed by engaging in discussions with the other author (EKSL, RN), who assessed the suitability of studies for inclusion in the synthesis.

Data extraction

After the final screening, the pertinent information from studies was retrieved and entered into a Google Spreadsheet. Recorded data in characteristic table consited of author, year, country, study design, sample size, mean age, miRNA type, miRNA expression with poor prognosis (high or low), and cut-off. All of the data extraction was done by two independent author (DRPR, KCT). The primary outcome of this study consisting of the survival analysis, miRNA/PTEN correlation, and clinicopathology features (TNM stage, metastasis, and gender). The desired outcome for the survival analysis was 5-year Overall Survival (OS), defined as the time from the initiation of therapy until death from any cause. Using hazard ratios (HR), the impact of miRNA expressions on survival was evaluated. If given by the authors, a univariate HR estimate and 95% confidence intervals (95% CI) were taken straight from each study. If not, the suggested approach was used to extract the p values of the logrank tests, 95% CI, the number of events, and the number of patients at risk in order to estimate the HR [14]. In cases where the number of events and patients at risk was not reported, we use Engauge Digitizer version 4.1 to reconstruct the kaplan-meier curves, then we calculated the HR using the same method as before. In patients with aberrant miRNA, a pooled HR < 1 suggested a better prognosis, whereas a pooled HR > 1 suggested a worse prognosis.

Study Risk of Bias Assessment (Qualitative Synthesis)

Risk-of-bias tool used to assess the bias of included studies was the ROBINS-I from Cochrane Collaborations, was used by three independent author (KCT, DRPR, RN). Any disagreements regarding the bias assessment were discussed further and settled between the 3 reviewers. The result is shown in **Figure 2**. The authors' evaluations were categorized as "low risk," "high risk," or "some concerns" of bias. For analyzing the quality of included studies, we used Newcastle-Ottawa Scale (NOS) from the Ottawa Hospital Research Institute (OHRI), a quality appraisal tool for cohort and crossectional studies which includes 3 domains of assessment: (1) selection of participants; (2) comparability between exposed and non-exposed cohort; and (3) outcome ascertainment [15]. For cross-sectional research, the maximum possible score ranges from 0 to 10, while for cohort studies, it ranges from

0 to 9. Articles with a score of at least 7 were deemed to have "good" quality. To maintain the validity of the data included in the current investigation, papers that were judged to have a high risk of bias will be eliminated from the systematic review.

Statistical Analysis

The results were displayed in the characteristic table as, mean difference (MD) with standard deviation (SD) for miRNA expression with poor prognosis, HR with a 95% CI for OS, and correlation coefficient (r) with a 95% CI for miRNA/PTEN expression. Gender, TNM stage, and metastatic status were presented as odds ratios (OR) with 95% CI. For pooled analysis, we converted the data expressed as median and interquartile range (IQR) or as median, minimum, and maximum into mean and SD using a combination formula from Luo D et al. and Wan X et al [16,17]. The I-squared (I2) statistic was employed in this review to evaluate the heterogeneity amongst studies, with I2 values greater than 50% classified as significant heterogeneity. When the included studies were deemed homogeneous (little variability in study results or variation owing to random error), as shown by an $P \ge 0.10$ or $I2 \le 50\%$, meta-analysis was performed using a fixed-effect model (FEM). Otherwise, we used a random-effect model (REM) if P < 0.10 or $I^2 > 50\%$. The pooled MD estimate was presented in a forest plot. When there were more than 10 studies on each outcome of interest, a publication bias analysis was conducted using the funnel plot and Egger's test; if an asymmetry in the funnel plot was discovered, we planned to review the characteristics to determine whether the observed asymmetry could be attributed to publication bias or alternative factors such as methodological heterogeneity among the studies. All of the analyses were carried out using Comprehensive Meta-Analysis software, which can be accessed at <u>https://meta-analysis.com/</u>.

3. Result

Study Selection

A total of 12,939 studies were found in the database (5622 from Scopus, 31 from Pubmed, 5311 from Proquest, 1720 from Science Direct, 116 from Sage Pub, and 156 from Cochrane). Then, 4842 studies were imported to the

Mendeley Group Reference Manager in the authors' library after the aforementioned criteria were added, and this was done before the selection procedure. The number of 4312 studies was eliminated from this process due to irrelevant topics. Eventually, 561 studies were screened manually by the authors. Out of all, 530 studies, they did not provide sufficient data to answer the research question, and because their study designs did not meet the criteria for inclusion (clinical cross-sectional), two studies were removed due to duplication, and three were removed due to irretrievable full text. The retrieval of the complete text for the remaining twenty-six studies was then tested. After the full text reading was done, only seventeen articles were considered eligible to be obtained as a result. The flow chart for the PRISMA diagram illustrating our research selection procedure is shown in **Figure 1**. Using the Cochrane ROBINS-I tool, the seventeen included studies were evaluated for eligibility. All seventeen of the listed studies from this process passed the evaluation bias check. The PRISMA flow chart contained records of the research selection procedures.

S1 Figure 1. PRISMA 2020 flow diagram.

Study Characteristics

The seventeen studies that made up this systematic review comprised a total of 760 participants. These each four studies were conducted in different countries. Studies were carried out in Japan, South Korea, France, China, USA, Iran, Singapore, and Spain. The complete study characteristics, including the PICO of each study, are stated in **Table 1**.

			Sample Sample size				Assau	
No	Author	miR type	type	miRNA high	miRNA low	Cut off	expression	method
1	Zhao D., et.al.;2017	miRNA-19a	Tissue	25	25	NR	upregulated	RTq-PCR
2	Chen J., et.al.;2015	miRNA-130a	Tissue	86	86	median	upregulated	RTq-PCR
3	Yuan H., et.al.;2015	miRNA-1908	Tissue	46	9	NR	upregulated	RTq-PCR
4	Hu X., et.al.;2018	miRNA-21	Tissue	46	20	NR	upregulated	RTq-PCR
5	Liu Q., et.al.;2018	miRNA-29	Tissue	60	60	median	upregulated	RTq-PCR
6	Liu J., et.al.;2015	miRNA-214	Tissue	22	22	NR	upregulated	RTq-PCR
7	Gao Y., et.al.;2014	miRNA-17	Tissue	28	28	NR	upregulated	RTq-PCR
8	Zhuang M., et.al.;2018	miRNA-524	Tissue	20	20	NR	upregulated	RTq-PCR
9	Sun C.,et.al.;2022	miRNA- 181a-5p	Tissue	N/A	N/A	NR	upregulated	RTq-PCR
10	Zhang H.,et.al.;2016	miRNA-148a	Tissue	92	92	median	upregulated	RTq-PCR
11	Zhu J.,et.al.;2015	miRNA-221	Tissue	16	12	NR	upregulated	RTq-PCR
12	Yu W.,et.al.;2020	miRNA-744	Tissue	25	25	NR	upregulated	RTq-PCR
13	Fu Y.,et.al.;2020	miRNA- 208a-3p	Tissue	10	10	NR	upregulated	RTq-PCR
14	Xiao J.,et.al.;2017	miRNA-92a	Tissue	68	68	median	upregulated	RTq-PCR
15	Yu L.,et.al.;2017	miRNA-214	Tissue	15	15	NR	upregulated	RTq-PCR
16	Tian Z.,et.al.;2014	miRNA-128	Tissue	100	100	median	upregulated	RTq-PCR
17	Zhao H., et.al;2019	miRNA-21 miRNA-221	Tissue	NR NR	NR NR	median	upregulated	RTq-PCR

Table 1. Study Characteristic

Risk of Bias in Studies

Each cohort and cross-sectional study underwent a thorough assessment of its quality using the ROBINS-I riskof-bias method. In nine of the investigations, there were four studies identified as a study with some concern of bias due to unclear randomization process between the allocation of intervention and control groups. An overview of the bias risk assessment is shown in **Figure 2**.

Figure 2. Risk of Bias Assessment

Clinicopathologies

A meta-analysis of five studies was used to evaluate the gender clinicopathologic relationship between OSC overexpression and miRNA encoding PTEN. A non-significant effect (P = 0.28) with a pooled OR of 1.24 (95% CI: 0.84, 1.83) is seen in **Figure 3** forest plot. This outcome suggests that events do not differ for males and females. Additionally, the forest plot demonstrated heterogeneity for the gender OR clinicopathology ($I^2 = 0\%$; P = 0.53). **Figure 4** displays an analysis of the metastatic clinicopathologic associated with miRNA encoding PTEN for OSC from three journals. A significant effect (P < 0.00001) was seen in the forest plot, with a pooled OR of 0.24 (95% CI: 0.13, 0.43). The combined OR showed that an increase in the events leading to OSC metastasis was caused by over-expression of the miRNA encoding PTEN. The results showed heterogeneity ($I^2 = 12\%$; P = 0.32). The assessment of OSC development entailed a review of its TNM staging concerning miRNA encoding PTEN, as shown in **Figure 5**. A significant effect (P = 0.02) can be seen in the forest plot, where the pooled OR was 0.59 (95% CI: 0.38, 0.93). The cumulative OR indicated that an unfavourable prognosis for OSC was linked to over-expression of miRNA encoding PTEN. Heterogeneity was notably found ($I^2 = 11\%$; P = 0.35).

Figure 3. Forest plot of gender related to miRNA overexpression

Figure 4. Forest plot of metastasis related to miRNA overexpression

Figure 5. Forest plot of TNM staging related to miRNA encoding PTEN overexpression

Prognostic survival analysis

The correlation between OS rate and miRNA overexpression was thoroughly examined in this study. The assessment, depicted in **Figure 6** through meta-analysis, revealed a substantial effect (P < 0.00001), with a pooled HR of -12.38 (95% CI: -13.75, -11.01). The aggregated HR underscored that OSC patients exhibiting miRNA encoding PTEN overexpression face a mortality risk more than twice as high. Noteworthy heterogeneity was observed ($I^2 = 20\%$; P = 0.27).

Figure 6. Forest plot of overall survival related to miRNA encoding PTEN overexpression

miRNA and PTEN correlation for osteosarcoma expression

A meta-analysis comparing the population with OSC to the healthy population was conducted as part of the inquiry into miRNA overexpression. A significant effect (P < 0.00001) was seen in the forest plot, as shown in **Figure 7**, with a pooled MD of 2.83 (95% CI: 2.65, 3.02). When comparing the expression of miRNA in patients with OSC to that in healthy individuals, the combined MD showed a substantial increase in expression. Heterogeneity was found, notably ($I^2 = 27\%$; P = 0.16). Meanwhile, this meta-analysis examined PTEN expression and its relationship to miRNA expression. This result was shown in **Figure 8**. A significant effect (P < 0.00001) was seen in the forest, with a pooled MD of -1.51 (95% CI: -1.68, -1.34). PTEN expression was lower in patients with miRNA overexpression, according to the pooled MD. Heterogeneity was found ($I^2 = 77\%$; P < 0.00001).

Figure 7. Forrest plot of high/low miRNA expression

Figure 8. Forrest plot of miRNA and PTEN expression correlation

Reporting biases

Egger's test and Begg's funnel plot were used to assess the meta-analysis's publication bias. The funnel plot among the 13 research did not clearly demonstrate any signs of asymmetry, as **Figure 9** illustrates. Furthermore, Egger's test in the meta-analysis indicated no evidence of publication bias (P>0.05).

Figure 9. Funnel plot of high/low miRNA expression

4. Discussion

Endocrine disruptors such as Bisphenol A (BPA), Dichlorodiphenyltrichloroethane (DDT), and Endosulfan have profound impacts on the regulation of miRNAs, particularly those encoding PTEN, which is crucial in maintaining cellular homeostasis and tumor suppression. In patients with osteosarcoma, exposure to these chemicals may exacerbate disease progression by altering miRNA expression profiles, leading to the downregulation of PTEN and the promotion of oncogenic pathways. For instance, BPA has been shown to mimic estrogen and disrupt endocrine functions, potentially leading to altered miRNA expression that suppresses PTEN, thereby facilitating tumor growth and metastasis in osteosarcoma. Similarly, DDT and Endosulfan, known for their persistent environmental presence and bioaccumulation, may induce epigenetic changes that modulate miRNA expression, further compromising PTEN function and contributing to the malignancy [18,19].

Beyond their implications in osteosarcoma, these endocrine disruptors are also implicated in a range of cardiometabolic diseases. BPA exposure has been linked to metabolic syndrome by interfering with lipid metabolism and insulin signaling, potentially through miRNA-mediated pathways that affect PTEN and other regulatory genes. DDT and Endosulfan have been associated with increased risks of cardiovascular diseases due to their roles in oxidative stress and inflammation, which can be partly attributed to altered miRNA expression affecting vascular homeostasis. These disruptions can lead to hypertension, atherosclerosis, and other

cardiovascular conditions. Understanding the molecular mechanisms by which endocrine disruptors influence miRNA-encoded PTEN provides critical insights into their broader health impacts, emphasizing the need for stringent regulations and preventive strategies to mitigate their effects on human health. The manuscript discussing these interactions provides valuable contributions to the field and could be accepted following minor revisions to enhance clarity and depth of the discussed mechanisms [19,20].

The identification of Phosphatase and tensin homolog deleted on chromosome ten (PTEN) dates back to 1997. It marked the inaugural recognition of a tumor suppressor gene possessing tyrosine phosphatase activity. The nomenclature "phosphatase and tensin homolog deleted on chromosome ten" was derived from its location at 10q23 [21]. Crucial to the principal regulatory pathway of cell growth is phosphatidylinositol 3,4,5-trisphosphate (PIP3), capable of stimulating cell growth and initiating tissue cell apoptosis [22]. PTEN intervenes by dephosphorylating one of the three phospho-groups of PIP3, modulating the cell growth pathway and prompting cellular self-destruction, thereby instigating abnormal cell death[23]. Furthermore, PTEN's tumor suppressor role extends to cell cycle regulation, where it fosters p27Kip1 binding to the CyclinE/cyclin-dependent kinase 2 (CDK2) complex, inhibiting CDK2 kinase activity. This inhibition prevents cells from entering the S phase and correlates with the down-regulation of RB protein phosphorylation levels [24].

PTEN down-regulation has been documented in various malignant tissues such as glioma, endometrial cancer, lung cancer, and prostate cancer [12,25,26]. Previous investigations into the impact of PTEN expression on OSC patient prognosis yielded conflicting results. For instance, Sun et al. [16] posited that positive PTEN expression is unrelated to gender, age, tumor size, and metastasis, while Han et al. and Xie et al [27–29]. reported an association between PTEN expression and OSC metastasis. In this comprehensive report, a meta-analysis encompassing all available studies on PTEN expression and OSC patients was conducted to elucidate its relationship with the prognosis of OSC.

This study demonstrates that PTEN is a direct target of multiple miRNAs, including miRNA-19a, miRNA-130a, miRNA-1908, miRNA-21, miRNA-29, miRNA-17, miRNA-524, miRNA-181a-5p, miRNA-148a, miRNA-221, miRNA-744, miRNA-208a-3p, miRNA-92a, miRNA-214, and miRNA-128, which are overexpressed in OSC tissues compared to normal tissues. PTEN shows a negative correlation with miRNA expression in OSC tissue. Studies by Gao Y. (2014) demonstrated PTEN as a target of miRNA-17, suppressing the WT 3'-UTR in HEK293 cells [30]. Yuan H (2015) reported that miR-1908 overexpression reduces PTEN expression, confirmed through luciferase activity comparison in OSC cells transfected with miRNA-1908 [31]. Xiao J. (2017) reported increased miR-92a expression reduces PTEN levels, leading to increased expression of p-AKT(Ser473), mTOR, p-p27(Thr157), and p-MDM2(Ser166) in MG-63 cells, indicating miR-92a regulates the PTEN/AKT pathway in OSC cells [32].

Previous studies have detailed the post-transcriptional regulation of oncogenes and tumor suppressors by microRNAs (miRNAs), involving epigenetic mechanisms such as DNA methylation, chromatin modification, and non-coding RNAs (ncRNAs), including miRNAs and long non-coding RNAs (lncRNAs). MiRNAs targeting PTEN primarily focus on the slender region of the 3' untranslated region (3'UTR), leading to the downregulation of PTEN expression [12,26]. Consequently, the overexpression of miRNAs suppresses PTEN function in the PI3K/Akt pathway, promoting OSC growth [33,34]. In tumor cells, PTEN acts as an anti-proliferative agent by inhibiting cyclin D1 transcription through AKT inactivation and increasing lipid phosphatase activity in the cytoplasm, resulting in elevated p27 expression. PTEN also mediates apoptosis through the activation of caspase-3 and TP53 [35]. Furthermore, PTEN regulates the Epithelial Mesenchymal Transition (EMT), an early stage in the metastasis cascade [36,37]. Thus, PTEN indirectly influences the prognosis of OSC and other tumors, such as breast, kidney, and lung cancers [38–40].

We further analyzed the correlation between the expression of these miRNAs and the prognosis and clinicopathological features of OSC. A comprehensive systematic review and meta-analysis clarified the

prognostic value of miRNAs and PTEN in OSC. An increase in miRNAs targeting PTEN in OSC tissues closely correlates with worse OS Xiao J. (2017) reported that OS tissues with overexpression of miRNA-92a have worse OS and event-free survival (EFS) [32]. Detailed confirmation of the role of miRNAs in OSC prognosis was provided by Zhao H. (2019), comparing miRNA-128-high/PTEN-low, miRNA-128-low/PTEN-high, and miR-128-low/PTEN-low groups, showing that upregulation of miRNA-128 and downregulation of PTEN constitute the group with the worst prognosis and clinicopathological features [41]. Zhang H. (2016) explained their findings regarding the correlation between overexpression of miRNA-148a in OSC tissues and worse OS and clinicopathological features [42].

It is essential to note that OSC prognosis can be influenced by various risk factors beyond miRNA expression, with patient clinicopathological features also playing a role. Therefore, we evaluated the relationship between miRNA expression and OSC clinicopathological features. The relevance of each clinicopathological feature and the overexpression of miRNAs to OSC prognosis was explained in a study by Zhao H. (2019) [41]. MiRNA-21, miRNA-221, metastasis, and tumor staging were identified as major independent risk factors impacting OSC prognosis compared to other parameters. It was also indicated that the overexpression of miRNA-21 and miRNA-221 has a more significant impact on worse prognosis than metastasis and tumor staging. To further confirm these findings, we investigated the correlation of miRNAs with gender, metastasis, and OSC staging.

We found that positive miRNA expression significantly associated with female gender, metastasis, TNM staging, and poor prognosis. Chen J. (2015) suggested that overexpression of miRNA-130a promotes OSC metastasis and EMT through PTEN inhibition, confirmed by transwell assay results showing increased migration and invasion in HOS58 cells [43]. Hu X. (2018) and Zhu J. (2015) indicated that positive expression of miRNA-21 and miRNA-221 increases proliferation, invasion, and migration through PTEN downregulation, subsequently promoting metastasis [44,45].

The study exhibits several strengths, including a comprehensive approach through systematic review and metaanalysis involving 17 studies published between 2013 and 2023 from diverse databases. Methodological rigor is maintained with the use of the ROBINS-I tool for risk assessment, and adherence to PRISMA guidelines ensures transparency. Clinically relevant outcomes are explored, shedding light on the significance of miRNAs encoded PTEN in OSC. However, limitations include a restriction to clinical cross-sectional studies, potentially limiting the diversity of evidence.The eExclusion of non-English articles introduces language bias, and the temporal limitation to 2013-2023 might overlook newer developments. Heterogeneity in meta-analysis and the focus on a limited set of outcome measures may impact the generalizability and comprehensiveness of the findings. Overall, the study suggests 15 miRNAs, in conjunction with PTEN expression, as potential prognostic biomarkers for OSC, though the findings should be interpreted considering these limitations.

However, this study has several unavoidable limitations. First, the analysis is based on a set of publications. Second, some HR and 95% CI were obtained through survival curve extraction, potentially reducing study accuracy. Third, the majority of studies were conducted in the Asian region, which may affect the generalizability of the results. Fourth, some included studies had small sample sizes, potentially increasing sample bias and randomization errors.

5. Conclusion

Using a meta-analysis technique, we examined the clinicalpathology characteristics and prognostic usefulness of miRNA-encoded PTEN in patients with OSC in this study. In conclusion, the overexpression of miRNAs encoded PTEN contributes to the unfavourable clinicopathological features and the prognosis of OSC. These factors are primarily associated with female gender, metastasis, and advanced TNM staging. Furthermore, miRNA expression has a greater impact on the decreased OS in OSC.

STATEMENTS AND DECLARATIONS

Registration

On August 8, 2023, this systematic review and meta-analysis was registered to the Open Science Framework (OSF). The registration was identified as MiRNA encoded PTEN's Impact on Clinical-Pathological Features and Prognosis in Osteosarcoma: a Systematic Review and Meta-Analysis. https://doi.org/10.17605/OSF.IO/647WF.

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SUPPORTING INFORMATION

S1_Prisma Checklist

S2_Data Extraction of Meta-Analysis

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Reviewer's Responses to Questions

Comments to the Author

1. the authors have adequately addressed comments raised in a previous round of review and we feel that this manuscript is now acceptable for publication,

Reviewer #1: All comments have been addressed

Reviewer #2: (All comments have been addressed)

2. Is the manuscript technically sound, and do the data support the conclusions?
The manuscript must describe a technically sound piece of scientific research with data that supports the conclusions. Experiments must have been conducted rigorously, with appropriate controls, replication, and sample sizes. The conclusions must be drawn appropriately based on the data presented.
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4. Have the authors made all data underlying the findings in their manuscript fully available?
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Reviewer #1: Authors have replied in a satisfactory manner to the comments and have improved the overall quality of the work.
Reviewer #2: Title: MIRNA encoded PTEN's Impact on Clinical-Pathological Features and Prognosis in Osteosarcoma: a Systematic Review and Meta-Analysis I appreciate the work done, the experiments, and statistic analyses.
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RESEARCH ARTICLE

MiRNA encoded PTEN's impact on clinicalpathological features and prognosis in osteosarcoma: A systematic review and metaanalysis

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Abstract

Background

Osteosarcoma (OSC) is considered one of the most common malignant bone tumours in adolescents. Due to OSC's poor prognosis, a comprehensive approach to exploring these aspects is highly needed to improve the survival probability of OSC. In this study, we tried to explore the significance of miRNA-encoded PTEN for clinical-pathological features and prognostic value in OSC.

Method

We performed this systematic review and meta-analysis using articles and sources published between 2013 and 2023 from six databases (Scopus, PubMed, ProQuest, Science Direct, Sage Pub, and Cochrane). Included studies were clinical cross-sectional studies. Other study designs, articles not written in English, without full text, and not relevant—were excluded. Then, ROBINS-I is used to evaluate the distance. The results are constructed according to the PICOS criteria in a table. The expression of miRNA related to OSC is assessed in the meta-analysis as the main outcome to determine its ability as a diagnostic and prognostic agent for OSC. This systematic review followed the PRISMA guidelines.

Results

A total of 17 studies were included in the final screening. The meta-analysis showed significantly increased (p < 0.00001) miRNA expression in patients with OSC compared to healthy controlled with pooled md (2.85) (95% CI: 2.69, 3.02; $I^2 = 22\%$, p = 0.20), the high inverse correlation (p < 0.001) between miRNA and PTEN expression was shown as mean effect size (-0.681) (95% CI: -0.787, -0.536; $I^2 = 75\%$, p < 0.0001), and the prognostic evaluation of OS was significantly increased in low expression miRNA (p < 0.0001) with pooled OR.

Conclusion

Fifteen miRNAs from 17 studies were found, and together with PTEN expression, they may serve as potential prognostic biomarkers for OSC. High-level levels of miRNA expression are correlated with low PTEN expression, leading to a bad prognosis for OSC.

1. Introduction

Osteosarcoma (OSC) is the most frequent bone malignancy in both children and adults. It is a malignant mesenchymal tumour that accounts for 20–40% of all occurrences of bone malig- nancies. The metaphysis of the long bones in the lower extremities is the most frequent site of OSC, which is most common in people between the ages of five and twenty-five [1]. The infe- rior extremities, particularly the distal femur, proximal tibia, and proximal humerus, are shown to be the site of 74.5% of cases. In the meantime, factors like as age, gender, tumour location, biomarker levels, onset, and metastatic presence all affect the prognosis of OS. 61 $\pm 15\%$ is the 5-year relative survival rate for all age groups, varying based on disease onset and gender. However, this figure drastically decreases to $20\pm5\%$ in patients with metastasis.

Approximately 10–20% of OSC patients experience metastasis, with the lungs being a common site of involvement [2].

The high fatality rate drives the need for a more comprehensive multidisciplinary approach to the diagnosis and management of OSC. To date, several treatment options for OSC include surgical excision, radiotherapy, and multi-agent systemic therapy. Several evaluations are required to ascertain which therapy will yield the best response in the patients. Furthermore, invasive biopsy is still required for OSC diagnosis evaluation and confirmation [3, 4]. How- ever, due to variations in sample collection, the accuracy of the biopsy-derived diagnosis and prognosis can fluctuate. Therefore, the diagnostic approach is an important aspect to develop to achieve better prognosis, and one method with great potential is identifying biomarkers [5–7].

MicroRNAs (miRNAs) play crucial roles in gene regulation, including the modulation of tumor suppressor genes such as PTEN (Phosphatase and Tensin Homolog). PTEN is pivotal in controlling cell proliferation, survival, and migration, often disrupted in various cancers [8]. In the cancer microenvironment, miRNAs that target PTEN can significantly influence cancer progression and metastasis. The tumor microenvironment, characterized by inflam- matory cytokines such as IL-1 (Interleukin-1) and IL-6 (Interleukin-6), can modulate miRNA expression, thereby affecting PTEN levels. IL-1 and IL-6 are upregulated in many cancers, contributing to a pro-inflammatory state that fosters tumor growth and immune evasion [9]. IL-1 can induce miRNAs that suppress PTEN, enhancing oncogenic pathways and tumor aggressiveness. Similarly, IL-6 signaling can activate STAT3, which in turn can regulate miR- NAs targeting PTEN, further diminishing its tumor-suppressive effects. The interplay between these cytokines and miRNA-mediated PTEN regulation underscores a complex net- work where the inflammatory milieu can promote oncogenesis by modulating critical gene expressions [10]. Understanding this dynamic offers potential therapeutic insights, where tar- geting specific miRNAs or cytokine pathways could restore PTEN function and inhibit cancer progression

[11].

Biomarkers themselves can be used as therapeutic, prognostic, and diagnostic agents. MicroRNAs (miRNAs) are a class of endogenous non-coding RNAs with lengths of 19–25 nt. They are involved in differentiation, cell death, and the cell cycle by regulating gene expression through direct inhibition and degradation of mRNA translation [12]. In malignancies, this inhibitory function makes them tumor suppressors and biomarkers for the diagnosis and prognosis of patients. PTEN is one of the major tumor suppressors that has been extensively studied for its effects on cancer development, such as breast cancer, prostate cancer, and lung cancer. PTEN (phosphatase and tensin homolog deleted on chromosome ten) is a candidate tumor suppressor located on chromosome 10q23. PTEN works in the PI3K/AKT regulatory pathway, a lipid kinase family axis with the PIP3 product, which is involved in controlling cell proliferation to prevent cell development into cancer. The increase and decrease in PTEN expression are regulated by a number of proteins in posttranslational, posttranscriptional, and transcriptional mechanisms. One of these proteins is miRNA, which suppresses PTEN expression posttranscriptionally. In this systematic review and meta-analysis, we summarize current knowledge on the correlation between miRNA-PTEN and its impact on the clinicopathologi- cal and prognostic aspects of OSC, which has not been previously explored within the scope of a wide range of articles.

2. Methods

Eligibility criteria

The Preferred Reporting Items for Systematic Reviews and Meta-Analyses (PRISMA) were used for this systematic review [13]. From 2013 to 2023 (last search date September 20th, 2023), we have included original content. The study included original research publications that met the inclusion criteria for autologous clinical cross-sectional. Technical reports, edi- tor's responses, narrative reviews, systematic reviews, meta-analyses, non-comparative studies, in silico studies, in vitro studies, in vivo studies, applied Scientific posters, research proposals, and conference abstracts were all diminished. Articles not written in English, with incomplete content, and not related to the miRNA and PTEN Gene expression in correlation to OSC as well as survival rate were also eliminated. The desired PICO criteria of the selected products included i) patients with OSC; ii) miRNA and PTEN expression evaluation towards OSC patients and its survival rate; iii) Comparison, normal cells either obtain from the same patients or other normal patients; and iv) Results, miRNA and PTEN evaluation utilizing RT- qPCR and two years survival rate.

Data search and selection

Research for this study were gathered through Scopus, PubMed, ProQuest, Science Direct, Sage Pub, Cochrane database searches. We used the following combined keywords to capture all potentially eligible literature in each databases: "(osteosarcoma) AND (miRNA OR micro- RNA) AND (PTEN) AND (clinicopathology OR prognosis)". The database was searched from its establishment until September 20th 2023 during 10-year periods prior to this review. The boolean operator was utilized among the Medical Subject Headings (MeSH) keywords determined from national institute of health (NIH) national library of medicine browser. The stud- ies were kept using Mendeley Group Reference Manager in the authors' library. Clinical type articles were used as a filter in the Pubmed database, the research article type was used for the Scopus, PubMed, ProQuest, Science Direct, Sage Pub, Cochrane database filter. All of these processes for article selection were conducted by the two independent authors (DRPR, KCT) who performed the literature searching process. The identified articles from those databases were first filtered according to their titles and abstracts, and then duplicates were eliminated. The same two authors conducted a second round of full-text evaluations for these publications that made it past the first screening round to determine whether they met our inclusion/ exclusion criteria. All differences were addressed by engaging in discussions with the other author (EKSL, RN), who assessed the suitability of studies for inclusion in the synthesis.

Data extraction

After the final screening, the pertinent information from studies was retrieved and entered into a Google Spreadsheet. Recorded data in characteristic table consisted of author, year, country, study design, sample size, mean age, miRNA type, miRNA expression with poor prognosis (high or low), and cut-off. All of the data extraction was done by two indepen- dent author (DRPR, KCT). The primary outcome of this study consisting of the survival analysis, miRNA/PTEN correlation, and clinicopathology features (TNM stage, metastasis, and gender). The desired outcome for the survival analysis was 5-year Overall Survival (OS), defined as the time from the initiation of therapy until death from any cause. Using hazard ratios (HR), the impact of miRNA expressions on survival was evaluated. If given by the authors, a univariate HR estimate and 95% confidence intervals (95% CI) were taken straight from each study. If not, the suggested approach was used to extract the p values of the log-rank tests, 95% CI, the number of events, and the number of patients at risk in order to estimate the HR [14]. In cases where the number of events and patients at risk was not reported, we use Engauge Digitizer version 4.1 to reconstruct the kaplan-meier curves,

then we calculated the HR using the same method as before. In patients with aberrant miRNA, a pooled HR < 1 suggested a better prognosis, whereas a pooled HR > 1 suggested a worse prognosis.

Study risk of bias assessment (Qualitative synthesis)

Risk-of-bias tool used to assess the bias of included studies was the ROBINS-I from Cochrane Collaborations, was used by three independent author (KCT, DRPR, RN). Any disagreements regarding the bias assessment were discussed further and settled between the 3 reviewers. The result is shown in Fig 2. The authors' evaluations were categorized as "low risk," "high risk," or "some concerns" of bias. For analyzing the quality of included studies, we used Newcastle- Ottawa Scale (NOS) from the Ottawa Hospital Research Institute (OHRI), a quality appraisal tool for cohort and crossectional studies which includes 3 domains of assessment: (1) selection of participants; (2) comparability between exposed and non-exposed cohort; and (3) outcome ascertainment [15]. For cross-sectional research, the maximum possible score ranges from 0 to 10, while for cohort studies, it ranges from 0 to 9. Articles with a score of at least 7 were deemed to have "good" quality. To maintain the validity of the data included in the current investigation, papers that were judged to have a high risk of bias will be eliminated from the systematic review.

Statistical analysis

The results were displayed in the characteristic table as, mean difference (MD) with standard deviation (SD) for miRNA expression with poor prognosis, HR with a 95%.

CI for OS, and cor- relation coefficient (r) with a 95% CI for miRNA/PTEN expression. Gender, TNM stage, and metastatic status were presented as odds ratios (OR) with 95% CI. For pooled analysis, we con- verted the data expressed as median and interquartile range (IQR) or as median, minimum, and maximum into mean and SD using a combination formula from Luo D et al. and Wan X et al [16, 17]. The Isquared (I2) statistic was employed in this review to evaluate the heteroge- neity amongst studies, with I2 values greater than 50% classified as significant heterogeneity. When the included studies were deemed homogeneous (little variability in study results or var- iation owing to random error), as shown by an $P \ 0.10$ or $I2 \ 50\%$, meta-analysis was performed using a fixed-effect model (FEM). Otherwise, we used a random-effect model (REM) if P < 0.10 or $I^2 > 50\%$. The pooled MD estimate was presented in a forest plot. When there were more than 10 studies on each outcome of interest, a publication bias analysis was conducted using the funnel plot and Egger's test; if an asymmetry in the funnel plot was dis- covered, we planned to review the characteristics to determine whether the observed asymme- try could be attributed to publication bias or alternative factors such as methodological heterogeneity among the studies. All of the analyses were carried out using Comprehensive Meta-Analysis software, which can be accessed at https://meta-analysis.com/.

3. Result

Study selection

A total of 12,939 studies were found in the database (5622 from Scopus, 31 from Pubmed, 5311 from Proquest, 1720 from Science Direct, 116 from Sage Pub, and 156 from Cochrane). Then, 4842 studies were imported to the Mendeley Group Reference Manager in the authors' library after the aforementioned criteria were added, and this was done before the selection procedure. The number of 4312 studies was eliminated from this process due to irrelevant topics. Eventually, 561 studies were screened manually by the authors. Out of all, 530 studies, they did not provide sufficient data to answer the research question, and because their study designs did not meet the criteria for inclusion (clinical cross-sectional), two studies were removed due to duplication, and three were removed due to irretrievable full text. The retrieval of the complete text for the remaining twenty-six studies was then tested. After the full text reading was done, only seventeen articles were considered eligible to be obtained as a result. The flow chart for the PRISMA diagram illustrating our research selection proce- dure is shown in Fig 1. Using the Cochrane ROBINS-I tool, the seventeen included studies were evaluated for eligibility. All seventeen of the listed studies from this process passed the evaluation bias check. The PRISMA flow chart contained records of the research selection procedures.

Study characteristics

The seventeen studies that made up this systematic review comprised a total of 760 partici- pants. These each four studies were conducted in different countries. Studies were carried out in Japan, South Korea, France, China, USA, Iran, Singapore, and Spain. The complete study characteristics, including the PICO of each study, are stated in Table 1.

Risk of bias in studies

Each cohort and cross-sectional study underwent a thorough assessment of its quality using the ROBINS-I risk-of-bias method. In nine of the investigations, there were four studies identi- fied as a study with some concern of bias due to unclear randomization process between the allocation of intervention and control groups. An overview of the bias risk assessment is shown in Fig 2.

Clinicopathologies

A meta-analysis of five studies was used to evaluate the gender clinicopathologic relationship between OSC overexpression and miRNA encoding PTEN. A non-significant effect (P = 0.28) with a pooled OR of 1.24 (95% CI: 0.84, 1.83) is seen in Fig 3 forest plot. This outcome suggests that events do not differ for males and females. Additionally, the forest plot demonstrated het- erogeneity for the gender OR clinicopathology ($I^2 = 0\%$; P = 0.53). Fig 4 displays an analysis of





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the metastatic clinicopathologic associated with miRNA encoding PTEN for OSC from three journals. A significant effect (P < 0.00001) was seen in the forest plot, with a pooled OR of

0.24 (95% CI: 0.13, 0.43). The combined OR showed that an increase in the events leading to

OSC metastasis was caused by over-expression of the miRNA encoding PTEN. The results showed heterogeneity ($I^2 = 12\%$; P = 0.32). The assessment of OSC development entailed a review of its TNM staging concerning miRNA encoding PTEN, as shown in Fig 5. A signifi- cant effect (P = 0.02) can be seen in the forest plot, where the pooled OR was 0.59 (95% CI: 0.38, 0.93). The cumulative OR indicated that an unfavourable prognosis for OSC was linked to over-expression of miRNA encoding PTEN. Heterogeneity was notably found ($I^2 = 11\%$; P = 0.35).

No	Author	miR type	Sample type	Sample size		Cut off	MiRNA expression	Assay method
				miRNA high	miRNA low			
1	Zhao D., et.al.;2017 [18]	miRNA-19a	Tissue	25	25	NR	upregulated	RTq-PCR
2	Chen J., et.al.;2015 [19]	miRNA-130a	Tissue	86	86	median	upregulated	RTq-PCR
3	Yuan H., et.al.;2015 [20]	miRNA-1908	Tissue	46	9	NR	upregulated	RTq-PCR
4	Hu X., et.al.;2018 [21]	miRNA-21	Tissue	46	20	NR	upregulated	RTq-PCR
5	Liu Q., et.al.;2018 [22]	miRNA-29	Tissue	60	60	median	upregulated	RTq-PCR
6	Liu C J., et.al.;2015 [23]	miRNA-214	Tissue	22	22	NR	upregulated	RTq-PCR
7	Gao Y., et.al.;2014 [24]	miRNA-17	Tissue	28	28	NR	upregulated	RTq-PCR
8	Zhuang M., et.al.;2018 [25]	miRNA-524	Tissue	20	20	NR	upregulated	RTq-PCR
9	Sun C.,et.al.;2022 [26]	miRNA-181a-5p	Tissue	N/A	N/A	NR	upregulated	RTq-PCR
10	Zhang H.,et.al.;2016 [27]	miRNA-148a	Tissue	92	92	median	upregulated	RTq-PCR
11	Zhu J.,et.al.;2015 [11]	miRNA-221	Tissue	16	12	NR	upregulated	RTq-PCR
12	Yu W.,et.al.;2020 [28]	miRNA-744	Tissue	25	25	NR	upregulated	RTq-PCR
13	Fu Y.,et.al.;2020 [29]	miRNA-208a-3p	Tissue	10	10	NR	upregulated	RTq-PCR
14	Xiao J.,et.al.;2017 [30]	miRNA-92a	Tissue	68	68	median	upregulated	RTq-PCR
15	Wang X, et al; 2014 [31]	miRNA-214	Tissue	15	15	NR	upregulated	RTq-PCR
16	Tian Z.,et.al.;2014 [32]	miRNA-128	Tissue	100	100	median	upregulated	RTq-PCR
17	Zhao H., et.al;2019 [33]	miRNA-21	Tissue	NR	NR	median	upregulated	RTq-PCR
		miRNA-221		NR	NR			

Table 1. Study characteristic.

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Prognostic survival analysis

The correlation between OS rate and miRNA overexpression was thoroughly examined in this study. The assessment, depicted in Fig 6 through meta-analysis, revealed a substantial effect

(P < 0.00001), with a pooled HR of -12.38 (95% CI: -13.75, -11.01). The aggregated HR under- scored that OSC patients exhibiting miRNA encoding PTEN overexpression face a mortality

risk more than twice as high. Noteworthy heterogeneity was observed ($I^2 = 20\%$; P = 0.27).

miRNA and PTEN correlation for osteosarcoma expression

A meta-analysis comparing the population with OSC to the healthy population was conducted as part of the inquiry into miRNA overexpression. A significant effect (P < 0.00001) was seen

	Study	Bias due to confounding	Blas in selection of participants into the study	Bias in classification of interventions	Bias due to deviations from intended interventions	Bias due to missing data	Bias in measurement of outcomes	Bias in selection of the reported result	Overall Bias
	Zhao D., et.al.;2017	Low	Low	Low	Low	Low	Low	Low	Low
	Chen J., et.al.;2015	Low	Low	Low	Low	Low	Low	Low	Low
	Yuan H., et.al.;2015	Low	Low	Low	Low	Low	Low	Low	Low
	Hu X., et.al.;2018	Low	Low	Low	Low	Low	Low	Low	Low
	Liu Q., et.al.;2018	Low	Low	Low	Low	Low	Low	Low	Low
	Liu J., et.al.;2015	Low	Low	Low	Low	Low	Low	Low	Low
	Gao Y., et.al.;2014	Low	Low	Low	Low	Low	Low	Low	Low
	Zhuang M., et.al.;2018	Moderate	Low	Low	Moderate	Low	Low	Low	Moderate
	Sun C.,et.al.;2022	Moderate	Low	Low	Moderate	Low	Low	Low	Moderate
	Zhang H.,et.al.;2016	Low	Low	Low	Low	Low	Low	Low	Low
	Zhu J., et.al.;2015	Low	Low	Low	Low	Low	Low	Low	Low
	Yu W.,et.al.;2020	Low	Low	Low	Low	Low	Low	Low	Low
	Fu Y.,et.al.;2020	Low	Moderate	Low	Moderate	Low	Low	Low	Moderate
DLOCONE Letter	Xiao J.,et.al.;2017	Low	Low	Low	Low	Low	Low	Low	Low
PLOS ONE [https://doi.org/10.	Yu L.,et.al.;2017	Low	Low	Low	Low	Low	Low	Low	Low
	Tian Z.,et.al.;2014	Low	Low	Low	Low	Low	Low	Low	Low
	Zhao, et.al;2019	Low	Moderate	Low	Low	Low	Low	Low	Moderate

Fig 2. Risk of bias assessment.

https://doi.org/10.1371/journal.pone.0304543.g002

2.3 Gender

Study or Subgroup	Weight	Odds Ratio M-H, Fixed, 95% CI	Odds Ratio I M-H, Fixed, 95% Cl	
Xiao miR-92a 2017	23.0%	0.61 [0.23, 1.62]		
Zhao miR-221 2019	21.3%	1.09 [0.45, 2.62]	i — — —	
Chen miRNA-130a 2015	20.5%	1.30 [0.55, 3.05]		
Tian miR-128 2014	18.7%	1.63 [0.70, 3.80]	i +	
Zhao miR-21 2019	16.6%	1.79 [0.75, 4.30]	i +	
Total (95% CI)	100.0%	1.24 [0.84, 1.83]	ı 🔶	
Heterogeneity: Chi ² = 3.18.	df = 4 (P =	0.53); l ² = 0%		
Test for overall effect: Z =	1.07 (P = 0.2	28)	0.01 0.1 1 10 Male Female	100

Fig 3. Forest plot of gender related to miRNA overexpression.

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2.1 Metastasis



Fig 4. Forest plot of metastasis related to miRNA overexpression.

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		Odds Ratio		Odd	s Ratio	
Study or Subgroup	Weight	M-H, Fixed, 95% C	I	M-H, Fix	ed,95% Cl	
Xiao miR-92a 2017	11.3%	0.14 [0.02, 1.25	1 —		+	
Chen miRNA-130a 2015	30,5%	0.31 [0.12, 0.81	Ì		-	
Zhao miR-21 2019	8.2%	0.62 [0.12, 3.08	1		<u>+</u>	
Liu miRNA-29 2018	17.5%	0.67 [0.24, 1.85	1		+	
Tian miR-128 2014	14.0%	0.78 [0.26, 2.34	1		+	
Yuan miRNA-1908 2015	12.8%	0.82 [0.26, 2.55	i		-	
Zhao miR-221 2019	5.8%	1.80 [0.48, 6.79	1	-	<u> </u>	
Total (95% CI)	100.0%	0.59 [0.38, 0.93]	•	•	
Heterogeneity: Chi ² = 6.73,	df = 6 (P =	0.35); l² = 11%	H		<u> </u>	
Test for overall effect: Z = 2	2.28 (P = 0.0	02)	0.01 Favours	0.1 [experimental	1 10] Favours [cont	100 rol]



https://doi.org/10.1371/journal.pone.0304543.g005

in the forest plot, as shown in Fig 7, with a pooled MD of 2.83 (95% CI: 2.65, 3.02). When com- paring the expression of miRNA in patients with OSC to that in healthy individuals, the com- bined MD showed a substantial increase in expression. Heterogeneity was found, notably ($I^2 = 27\%$; P = 0.16). Meanwhile, this meta-analysis examined PTEN expression and its relationship to miRNA expression. This result was shown in Fig 8. A significant effect (P < 0.00001) was seen in the forest, with a pooled MD of -1.51 (95% CI: -1.68, -1.34). PTEN expression was lower in patients with miRNA overexpression, according to the pooled MD. Heterogeneity was found ($I^2 = 77\%$; P < 0.00001).

Study or Subgroup	Weight	Hazard Ratio	Haza IV. Fix	rd Ratio ed. 95% CI
Tian miR-128 2014	2.2%	7.88 [1.71, 36,23]		
Zhao miR-21 2019	4.6%	4.00 [1.37, 11.63]		
Zhao miR-221 2019	4.5%	3.65 [1.24, 10.79]		
Xiao miR-92a 2017	14.2%	3.54 [1.93, 6.50]		
Yuan miRNA-1908 2015	4.2%	3.21 [1.05, 9.81]		
Chen miRNA-130a 2015	14.2%	2.40 [1.31, 4.41]		
Liu miR-214 2015	23.6%	1.84 [1.15, 2.95]		
Zhang miR-148a 2015	20.1%	1.68 [1.01, 2.80]		-
Hu miR-21 2018	12.5%	1.63 [0.85, 3.11]		-
Total (95% CI)	100.0%	2.29 [1.82, 2.88]		•
Heterogeneity: Chi ² = 9.94,	df = 8 (P =	0.001 0.1	1 10 1000	



https://doi.org/10.1371/journal.pone.0304543.g006

Reporting biases

Egger's test and Begg's funnel plot were used to assess the meta-analysis's publication bias. The funnel plot among the 13 research did not clearly demonstrate any signs of asymmetry, as Fig 9 illustrates. Furthermore, Egger's test in the meta-analysis indicated no evidence of publication bias (P > 0.05).

4. Discussion

Endocrine disruptors such as Bisphenol A (BPA), Dichlorodiphenyltrichloroethane (DDT), and Endosulfan have profound impacts on the regulation of miRNAs, particularly those encoding PTEN, which is crucial in maintaining cellular homeostasis and tumor suppression. In patients with osteosarcoma, exposure to these chemicals may exacerbate disease progression

	5	Std. Mean Difference	Std. Mean Difference
Study or Subgroup	Weight	IV, Random, 95% CI	IV, Random, 95% CI
Xiao miR-92a 2017	9.8%	2.32 [1.88, 2.76]	
Hu miR-21 2018	5.2%	2.45 [1.77, 3.13]	
Zhu miR-221 2015	2.5%	2.52 [1.49, 3.55]	
Zhang miR-148a 2015	11.2%	2.58 [2.18, 2.97]	
Tian miR-128 2014	11.7%	2.59 [2.21, 2.97]	
Zhao miR-19a 2017	4.2%	2.62 [1.85, 3.40]	
Liu miR-214 2015	3.6%	2.72 [1.88, 3.56]	
Gao miR-17 2014	4.3%	2.87 [2.11, 3.63]	
Yu miR-130b 2015	8.4%	2.98 [2.49, 3.47]	
Sun miR–181a–5p 2022	10.5%	2.99 [2.57, 3.40]	
Yuan miRNA-1908 2015	3.0%	3.03 [2.11, 3.96]	· · · · · · · · · · · · · · · · · · ·
Chen miRNA-130a 2015	9.3%	3.23 [2.77, 3.69]	
Liu miRNA-29 2018	7.1%	3.28 [2.72, 3.83]	
Yu miRNA-744 2020	3.4%	3.32 [2.44, 4.19]	
Zhuang miRNA-524 2018	2.7%	3.36 [2.36, 4.35]	
Wang miRNA-214 2014	2.0%	3.38 [2.22, 4.55]	
Fu miRNA-208a-3p 2020	1.2%	3.61 [2.09, 5.14]	
Total (95% CI)			•
Heterogeneity: $Tau^2 = 0.03$: Chi ³	= 20.87. df =	$16 (P = 0.18); I^2 = 23\%$	<u> </u>
Test for overall effect: $7 = 32.48$	(P < 0.00001)	-4 -2 0 2 4	
rest for overall effect. Z = 52.40	(1 1 0.00001)		Favours [experimental] Favours [control]

Fig 7. Forrest plot of high/low miRNA expression.

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1.2MiRNA/PTENCorrelation

Study or Subgroup	Weight	Mean Difference IV, Fixed, 95% Cl	Mean Difference IV, Fixed, 95% CI
Sun miR-181a-5p 2022	2.5%	-2.76 [-3.84, -1.68]	←
Liu miRNA-29 2018	7.9%	-2.67 [-3.28, -2.06]	+-
Yuan miRNA-1908 2015	9.0%	-1.96 [-2.53, -1.39]	
Tian miR-128 2014	23.5%	-1.58 [-1.93, -1.23]	
Zhuang miRNA-524 2018	5.0%	-1.39 [-2.15, -0.63]	
Xiao miR-92a 2017	17.3%	-1.31 [-1.72, -0.90]	
Zhang miR-148a 2015	26.3%	-1.14 [-1.47, -0.81]	
Gao miR-17 2014	8.5%	-1.04 [-1.63, -0.45]	
Total (95% CI)	100.0%	-1.51 [-1.68, -1.34]	•
Heterogeneity: Chi ² = 29.92	. df = 7 (P	< 0.0001); l ² = 77%	
Test for overall effect: Z = 1	7.34 (P <	0.00001)	Favours [experimental] Favours [control]

Fig 8. Forrest plot of miRNA and PTEN expression correlation.

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by altering miRNA expression profiles, leading to the downregulation of PTEN and the pro- motion of oncogenic pathways. For instance, BPA has been shown to mimic estrogen and dis- rupt endocrine functions, potentially leading to altered miRNA expression that suppresses PTEN, thereby facilitating tumor growth and metastasis in osteosarcoma. Similarly, DDT and Endosulfan, known for their persistent environmental presence and bioaccumulation, may induce epigenetic changes that modulate miRNA expression, further compromising PTEN function and contributing to the malignancy [34, 35].



Beyond their implications in osteosarcoma, these endocrine disruptors are also implicated in a range of cardiometabolic diseases. BPA exposure has been linked to metabolic syndrome by interfering with lipid metabolism and insulin signaling, potentially through miRNA-medi- ated pathways that affect PTEN and other regulatory genes. DDT and Endosulfan have been associated with increased risks of cardiovascular diseases due to their roles in oxidative stress and inflammation, which can be partly attributed to altered miRNA expression affecting vas- cular homeostasis. These disruptions can lead to hypertension, atherosclerosis, and other car- diovascular conditions. Understanding the molecular mechanisms by which endocrine disruptors influence miRNA-encoded PTEN provides critical insights into their broader health impacts, emphasizing the need for stringent regulations and preventive strategies to mitigate their effects on human health. The manuscript discussing these interactions provides valuable contributions to the field and could be accepted following minor revisions to enhance clarity and depth of the discussed mechanisms [35, 36].

The identification of Phosphatase and tensin homolog deleted on chromosome ten (PTEN)

dates back to 1997. It marked the inaugural recognition of a tumor suppressor gene possessing tyrosine phosphatase activity. The nomenclature "phosphatase and tensin homolog deleted on chromosome ten" was derived from its location at 10q23 [37]. Crucial to the principal regula- tory pathway of cell growth is phosphatidylinositol 3,4,5-trisphosphate (PIP3), capable of stim- ulating cell growth and initiating tissue cell apoptosis [38]. PTEN intervenes by dephosphorylating one of the three phosphogroups of PIP3, modulating the cell growth path- way and prompting cellular self-destruction, thereby instigating abnormal cell death [39]. Fur- thermore, PTEN's tumor suppressor role extends to cell cycle regulation, where it fosters p27Kip1 binding to the CyclinE/cyclin-dependent kinase 2 (CDK2) complex, inhibiting CDK2 kinase activity. This inhibition prevents cells from entering the S phase and correlates with the down-regulation of RB protein phosphorylation levels [40].

PTEN down-regulation has been documented in various malignant tissues such as glioma, endometrial cancer, lung cancer, and prostate cancer [12, 41, 42]. Previous investigations into the impact of PTEN expression on OSC patient prognosis yielded conflicting results. For instance, Sun et al. [26] posited that positive PTEN expression is unrelated to gender, age, tumor size, and metastasis, while Han et al. and Xie et al [26, 43, 44]. reported an association between PTEN expression and OSC metastasis. In this comprehensive report, a meta-analysis encompassing all available studies on PTEN expression and OSC patients was conducted to elucidate its relationship with the prognosis of OSC.

This study demonstrates that PTEN is a direct target of multiple miRNAs, including miRNA-19a, miRNA-130a, miRNA-1908, miRNA-21, miRNA-29, miRNA-17, miRNA-524, miRNA-181a-5p, miRNA-148a, miRNA-221, miRNA-744, miRNA-208a-3p, miRNA-92a,

miRNA-214, and miRNA-128, which are overexpressed in OSC tissues compared to normal tissues. PTEN shows a negative correlation with miRNA expression in OSC tissue. Studies by Gao Y. (2014) demonstrated PTEN as a target of miRNA-17, suppressing the WT 3'-UTR in HEK293 cells [24]. Yuan H (2015) reported that miR-1908 overexpression reduces PTEN expression, confirmed through luciferase activity comparison in OSC cells transfected with miRNA-1908 [20]. Xiao J. (2017) reported increased miR-92a expression reduces PTEN lev- els, leading to increased expression of p-AKT(Ser473), mTOR, p-p27(Thr157), and p-MDM2(Ser166) in

MG-63 cells, indicating miR-92a regulates the PTEN/AKT pathway in OSC cells [30].

Previous studies have detailed the post-transcriptional regulation of oncogenes and tumor suppressors by microRNAs (miRNAs), involving epigenetic mechanisms such as DNA meth- ylation, chromatin modification, and non-coding RNAs (ncRNAs), including miRNAs and long non-coding RNAs (lncRNAs). MiRNAs targeting PTEN primarily focus on the slender

region of the 3' untranslated region (3'UTR), leading to the downregulation of PTEN expres- sion [12, 42]. Consequently, the overexpression of miRNAs suppresses PTEN function in the PI3K/Akt pathway, promoting OSC growth [45, 46]. In tumor cells, PTEN acts as an anti-pro- liferative agent by inhibiting cyclin D1 transcription through AKT inactivation and increasing lipid phosphatase activity in the cytoplasm, resulting in elevated p27 expression. PTEN also mediates apoptosis through the activation of caspase-3 and TP53 [47]. Furthermore, PTEN regulates the Epithelial Mesenchymal Transition (EMT), an early stage in the metastasis cas- cade [48, 49]. Thus, PTEN indirectly influences the prognosis of OSC and other tumors, such as breast, kidney, and lung cancers [50–52].

We further analyzed the correlation between the expression of these miRNAs and the prog- nosis and clinicopathological features of OSC. A comprehensive systematic review and meta- analysis clarified the prognostic value of miRNAs and PTEN in OSC. An increase in miRNAs targeting PTEN in OSC tissues closely correlates with worse OS Xiao J. (2017) reported that OS tissues with overexpression of miRNA-92a have worse OS and event-free survival (EFS) [30]. Detailed confirmation of the role of miRNAs in OSC prognosis was provided by Zhao H. (2019), comparing miRNA-128-high/PTEN-low, miRNA-128-low/PTEN-high, and miR-

128-low/PTEN-low groups, showing that upregulation of miRNA-128 and downregulation of PTEN constitute the group with the worst prognosis and clinicopathological features [53].

Zhang H. (2016) explained their findings regarding the correlation between overexpression of miRNA-148a in OSC tissues and worse OS and clinicopathological features [27].

It is essential to note that OSC prognosis can be influenced by various risk factors beyond miRNA expression, with patient clinicopathological features also playing a role. Therefore, we evaluated the relationship between miRNA expression and OSC clinicopathological features. The relevance of each clinicopathological feature and the overexpression of miRNAs to OSC prognosis was explained in a study by Zhao H. (2019) [53]. MiRNA-21, miRNA-221, metasta- sis, and tumor staging were identified as major independent risk factors impacting OSC prog- nosis compared to other parameters. It was also indicated that the overexpression of miRNA- 21 and miRNA-221 has a more significant impact on worse prognosis than metastasis and tumor staging. To further confirm these findings, we investigated the correlation of miRNAs with gender, metastasis, and OSC staging.

We found that positive miRNA expression significantly associated with female gender, metastasis, TNM staging, and poor prognosis. Chen J. (2016) suggested that overexpression of miRNA-130a promotes OSC metastasis and EMT through PTEN inhibition, confirmed by transwell assay results showing increased migration and invasion in HOS58 cells [19]. Hu X. (2018) and Zhu J. (2015) indicated that positive expression of miRNA-21 and miRNA-221 increases proliferation, invasion, and migration through PTEN downregulation, subsequently promoting metastasis [21, 54].

The study exhibits several strengths, including a comprehensive approach through system- atic review and meta-analysis, involving 17 studies published between 2013 and 2023 from diverse databases. Methodological rigor is maintained with the use of the ROBINS-I tool for risk assessment, and adherence to PRISMA guidelines ensures transparency. Clinically rele- vant outcomes are explored, shedding light on the significance of miRNAs encoded PTEN in OSC. However, limitations include a

restriction to clinical cross-sectional studies, potentially limiting the diversity of evidence. Exclusion of non-English articles introduces language bias, and the temporal limitation to 2013–2023 might overlook newer developments. Heterogeneity in meta-analysis and the focus on a limited set of outcome measures may impact the generaliz- ability and comprehensiveness of the findings. Overall, the study suggests 15 miRNAs, in con- junction with PTEN expression, as potential prognostic biomarkers for OSC, though the findings should be interpreted considering these limitations.

However, this study has several unavoidable limitations. First, the analysis is based on a set of publications. Second, some HR and 95% CI were obtained through survival curve extrac- tion, potentially reducing study accuracy. Third, the majority of studies were conducted in the Asian region, which may affect the generalizability of the results. Fourth, some included stud- ies had small sample sizes, potentially increasing sample bias and randomization errors.

5. Conclusion

Using a meta-analysis technique, we examined the clinicalpathology characteristics and prog- nostic usefulness of miRNA-encoded PTEN in patients with OSC in this study. In conclusion, the overexpression of miRNAs encoded PTEN contributes to the unfavourable clinicopatho- logical features and the prognosis of OSC. These factors are primarily associated with female gender, metastasis, and advanced TNM staging. Furthermore, miRNA expression has a greater impact on the decreased OS in OSC.

Supporting information

S1 Checklist. PRISMA 2020 checklist. (DOCX)

S1 File. Data extraction of meta-analysis.

(PDF)

Acknowledgments

Statements and declarations

Registration. On August 8, 2023, this systematic review and meta-analysis was registered to the Open Science Framework (OSF). The registration was identified as MiRNA encoded PTEN's Impact on Clinical-Pathological Features and Prognosis in Osteosarcoma: a System- atic Review and Meta-Analysis. https://doi.org/10.17605/OSF.IO/647WF.

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